

Bayesian Statistics with R-INLA

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Getting INLA

Implementing the INLA algorithm

How to use INLA

Simple example

Add random effects

Prediction

Smoothing binary time series

Disease Mapping

Changing the prior

Repeated Poisson counts

Control statements

Getting INLA Implementing the INLA algorithm How to use INLA Simple example Add random effects Prediction

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Model Comparison

Getting INLA	Implementing the INLA algorithm	How to use INLA	Simple example	Add random effects	Predictions
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What have we learned in the morning...

- What is a LGM
- Which kind of models are amenable to INLA
- How does INLA work....

What have we learned in the morning...

- What is a LGM
- Which kind of models are amenable to INLA
- How does INLA work....
- ..you have even implemented it yourself! :-)

Good News!

All the theory we have seen is wrapped up in the R-package
INLA which is easy to use.

Getting INLA

Getting INLA

- The web page www.r-inla.org contains source-code, worked-through examples, reports and instructions for installing the package.

Getting INLA

- The R-package INLA works on Linux, Windows and Mac and can be installed within R by

```
# stable version
install.packages("INLA",
  repos=c(getOption("repos"),
    INLA="https://inla.r-inla-download.org/R/stable"),
  dep=TRUE)

# devel version
install.packages("INLA",
  repos=c(getOption("repos"),
    INLA="https://inla.r-inla-download.org/R/testing"),
  dep=TRUE)
```

and then upgraded in R as:

```
inla.upgrade(testing = TRUE)
```

****NB** You need R version 4.1 or newer!!**

INLA runs in parallel!

INLA can run in parallel for faster computations with large models.

It uses the **PARDISO 7.2 Solver Project** and you need to get a license to use it!

```
library(INLA)  
inla.pardiso()
```

..and follow the instruction there!

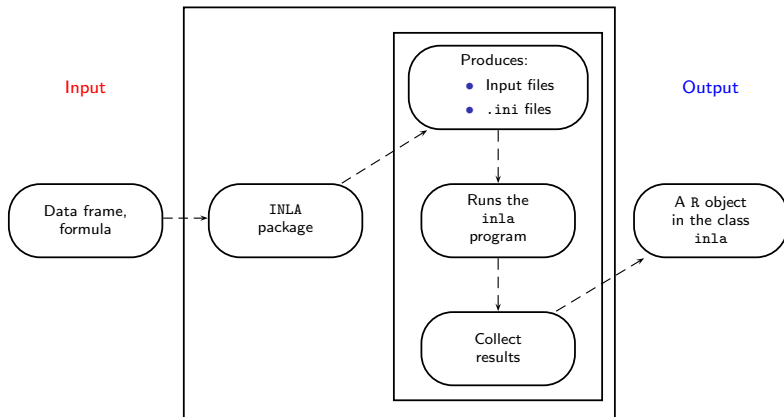
Which INLA version do I have?

```
inla.version()
```

```
## R-INLA version .....: 22.02.16-2
## Date .....: Wed Feb 16 02:22:38 PM +03 2
## Maintainers .....: Havard Rue <hrue@r-inla.org>
##                  : Finn Lindgren <finn.lindgren@r-inla.org>
##                  : Elias Teixeira Krainski <elias.krainski@r-inla.org>
## Main web-page .....: www.r-inla.org
## Download-page .....: inla.r-inla-download.org
## Repository .....: github.com/hrue/r-inla
## Email support .....: help@r-inla.org
##                  : r-inla-discussion-group@googlegroups.com
```

Implementing the INLA algorithm

The INLA package for R



What happens in the black box?

The implementation of the INLA method consists of three parts:

- **GMRFLib-Library:** A library for GMRFs written in C
- **inla-program:** The implementation of INLA written in C
- **INLA package for R:** An R-interface to the inla-program

The first two are *not* particularly user-friendly. They are used in the background by the INLA package.

Implementing INLA

All procedures required to perform INLA need to be carefully implemented to achieve a good speed; easier to implement a slow version of INLA.

Implementing INLA

All procedures required to perform INLA need to be carefully implemented to achieve a good speed; easier to implement a slow version of INLA.

- **The GMRFLib-library**
 - Basic library written in C, user friendly for programmers

Implementing INLA

All procedures required to perform INLA need to be carefully implemented to achieve a good speed; easier to implement a slow version of INLA.

- **The GMRFLib-library**
- **The inla-program**
 - Define *latent Gaussian models* and interface with the GMRFLib-library
 - Avoids the need for C-programming
 - Models are defined using `.ini`-files
 - Requires to write input files in a special format
 - `inla-program` write all the results (E/Var/marginals) to files

Implementing INLA

All procedures required to perform INLA need to be carefully implemented to achieve a good speed; easier to implement a slow version of INLA.

- **The GMRFLib-library**
- **The inla-program**
- **The INLA package for R**
 - R-interface to the inla-program. (That's why its not on CRAN.)
 - Convert **formula**-statements into **.ini**-files definitions
 - It also does much more (for example for survival models or when using **inlabru**)

How to use INLA

How to use INLA

There are essentially four parts to an INLA-program:

1. **Data organisation**: Make an object to store response, covariates,

```
data = data.frame(y = y, x = x)
```

2. **Use the 'formula'-notation** to specify the model (similar to lm and glm functions)

```
formula = y~x
```

3. **Call the 'inla'-program**

```
res = inla(formula, data=data, family="gaussian")
```

4. **Extract posterior information**, e.g. for a first overview use

```
summary(res)
```

Data organization

The responses and covariates are collected in a **list or data frame**. Assume response y , covariates x_1 and x_2 , and time index t . Then they can be organized with:

Option 1

```
data = list(y = y, x1 = x1, x2 = x2, t = t)
```

Option 2

```
data = data.frame(y = y, x1 = x1, x2 = x2, t = t)
```

formula: specifying the linear predictor

The model is specified through a ‘formula’ similar to `glm`:

```
formula = y ~ x1 + x2 + f(t, ...)
```

- `y` is the name of the response in the `data` object
- The fixed effects are given i.i.d. Gaussian priors
- The `f()` function specifies random effects (e.g. temporal, spatial, smooth effect of covariates and Besag model)
- Use `-1` in the formula if you don't want an automatic intercept

The inla() function

```
result = inla(
  # Description of linear predictor
  formula,
  # Likelihood
  family = "gaussian",
  # List or data frame with response,
  # covariates, etc.
  data = data,
  ## This is all that is needed for a basic call

  ## # check what happens
  verbose = TRUE,
  # ,..., there are also some "control statements"
  # to customize things
  # This you need if you later want to sample from the
  # fitted model
  control.compute=list(config = TRUE)
)
```


Likelihood functions

- gaussian
- T
- poisson
- nbinoomial
- binomial
- exponential
- weibull
- gev
- coxph

For a complete list type

```
names(inla.models())$likelihood)
```

Posterior inference

Main functions:

```
# look at a first summary
summary(result)
# plot the main results
# (does not use ggplot...)
plot(result)
# rerun the model to get better
# estimate of the hyperparameters
result2 = inla.hyperpar(result)
# sample from the fitted model
# this can be very useful sometimes!
sample = inla.posterior.sample(results)
```

Simple example

Example: Simple linear regression

- **Stage 1:** Gaussian likelihood

$$y_i | \eta_i \sim \mathcal{N}(\eta_i, \sigma^2)$$

- **Stage 2:** Covariates are connected to likelihood by

$$\eta_i = \beta_0 + \beta_1 x_i$$

- **Stage 3:** σ^2 : variance of observation noise

Example: Simple linear regression

```
# Generate data
```

```
x = runif(10)
```

```
y = 1 + 2*x + rnorm(n = 100, sd = 0.1)
```

```
# Run inla
```

```
formula = y ~ 1 + x
```

```
result = inla(formula,  
              data = data.frame(x = x, y = y),  
              family = "gaussian")
```

Organization of the inla-object

```
names(result)
```

```
## [1] "names.fixed"           "summary.fixed"
## [3] "marginals.fixed"       "summary.lincomb"
## [5] "marginals.lincomb"     "size.lincomb"
## [7] "summary.lincomb.derived" "marginals.lincomb.derived"
## [9] "size.lincomb.derived"  "mlik"
## [11] "cpo"                   "po"
## [13] "waic"                   "model.random"
## [15] "summary.random"        "marginals.random"
## [17] "size.random"           "summary.linear.predictor"
## [19] "marginals.linear.predictor" "summary.fitted.values"
## [21] "marginals.fitted.values" "size.linear.predictor"
## [23] "summary.hyperpar"       "marginals.hyperpar"
## [25] "internal.summary.hyperpar" "internal.marginals.hyperpar"
## [27] "offset.linear.predictor" "model.spde2.blc"
## [29] "summary.spde2.blc"      "marginals.spde2.blc"
## [31] "size.spde2.blc"         "model.spde3.blc"
## [33] "summary.spde3.blc"      "marginals.spde3.blc"
## [35] "size.spde3.blc"        "logfile"
## [37] "misc"                   "dic"
## [39] "mode"                   "joint.hyper"
## [41] "nhyper"                 "version"
## [43] "Q"                      "graph"
## [45] "ok"                     "cpu.used"
## [47] "all.hyper"              ".args"
## [49] "call"                   "model.matrix"
```

Organization of the inla-object

You can find summary information in

```
## [1] "summary.fixed"           "summary.lincomb"
## [3] "summary.lincomb.derived" "summary.random"
## [5] "summary.linear.predictor" "summary.fitted.values"
## [7] "summary.hyperpar"       "internal.summary.hyperpar"
## [9] "summary.spde2.blc"      "summary.spde3.blc"
```

for example

```
result$summary.fixed
```

```
##              mean              sd 0.025quant 0.5quant 0.975quant      mode
## (Intercept) 1.002781 0.01981293  0.9638022 1.002780  1.041726 1.002781
## x           1.974175 0.03346575  1.9083371 1.974174  2.039957 1.974175
##              kld
## (Intercept) 3.086520e-06
## x           3.086389e-06
```

Organization of the `inla`-object

You can find estimated posterior marginals in

```
## [1] "marginals.fixed"           "marginals.lincomb"
## [3] "marginals.lincomb.derived" "marginals.random"
## [5] "marginals.linear.predictor" "marginals.fitted.values"
## [7] "marginals.hyperpar"       "internal.marginals.hyperpar"
## [9] "marginals.spde2.blc"      "marginals.spde3.blc"
```

Each object is thereby a list. Get the marginal for intercept:

```
head(result$marginals.fixed[[1]])
```

```
##           x           y
## [1,] 0.8043030 2.146677e-17
## [2,] 0.8439985 3.450320e-11
## [3,] 0.8836940 1.944195e-06
## [4,] 0.8941229 2.290543e-05
## [5,] 0.9035418 1.840298e-04
## [6,] 0.9043623 2.192137e-04
```


Organization of the inla-object

Further general information

formula used

```
result$.args$formula
```

```
## y ~ 1 + x
```

```
## NULL
```

data used

```
result$.args$data[1:3,]
```

```
##           x           y
```

```
## 1 0.8051767 2.617140
```

```
## 2 0.7827668 2.542227
```

```
## 3 0.1807834 1.157771
```

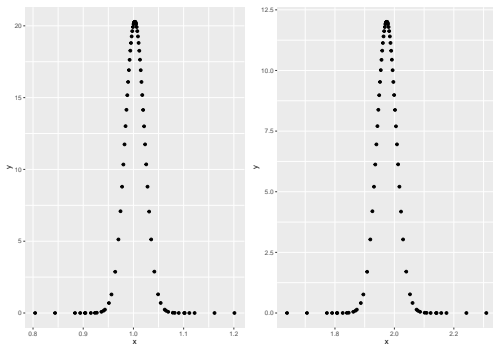
log-file including information of INLA approximations

```
result$logfile
```

Marginal posterior densities

The marginal posterior densities are stored as a matrices with x - and y -values

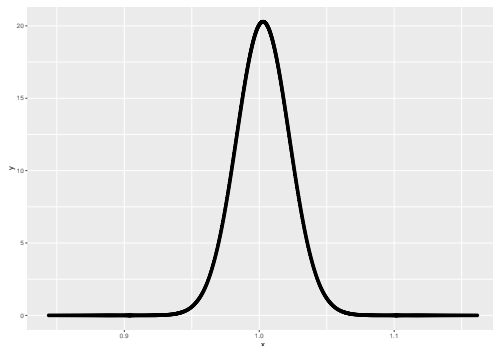
```
intercept = data.frame(result$marginals.fixed$`Intercept`)
x = data.frame(result$marginals.fixed$x)
p1 = ggplot(data = intercept) + geom_point(aes(x,y))
p2 = ggplot(data = x) + geom_point(aes(x,y))
p1+p2
```



Marginal posterior densities

The rough shape can be interpolated to higher resolution using the `inla.smarginal()` function:

```
smoother_dens = data.frame(inla.smarginal(intercept))
ggplot(data = smoother_dens) + geom_point(aes(x,y))
```



Marginal posterior densities

Manipulation of the computed posterior marginals is possible through the `inla.*marginal()` functions:

```
# compute the 0.05 quantile
```

```
inla.qmarginal(0.05, intercept)
```

```
## [1] 0.9701406
```

```
# Distribution function
```

```
inla.pmarginal(0.975, intercept)
```

```
## [1] 0.08047574
```

```
# Density function
```

```
inla.dmarginal(1, intercept)
```

```
## [1] 20.07664
```

```
# Generate realizations
```

```
inla.rmarginal(4, intercept)
```

```
## [1] 0.9969020 1.0146015 1.0341315 0.9967547
```

Other `inla.*marginal()` functions.

Function Name	Usage
<code>inla.dmarginal(x, marginal, ...)</code>	Density at a vector of evaluation points x
<code>inla.pmarginal(q, marginal, ...)</code>	Distribution function at a vector of quantiles q
<code>inla.qmarginal(p, marginal, ...)</code>	Quantile function at a vector of probabilities p .
<code>inla.rmarginal(n, marginal)</code>	Generate n random deviates
<code>inla.hpdmarginal(p, marginal, ...)</code>	Compute the highest posterior density interval at level p
<code>inla.emarginal(fun, marginal, ...)</code>	Compute the expected value of the marginal assuming the transformation given by <code>fun</code>
<code>inla.mmarginal(marginal)</code>	Compute the mode
<code>inla.smarginal(marginal, ...)</code>	Smoothed density in form of a list of length two. The first entry contains the x-values, the second entry includes the interpolated y-values
<code>inla.tmarginal(fun, marginal, ...)</code>	Transform the marginal using the function <code>fun</code> .
<code>inla.zmarginal(marginal)</code>	Summary statistics for the marginal

Add random effects

Add random effects

```
f(name, model="...", hyper=...,
    constr=FALSE, cyclic=FALSE, ...)
```

- `name` – the index of the effect (each f-function needs its own!)
- `model` – the type of latent model. E.g. iid, rw2, ar1, besag, and so on
- `hyper` – specify the prior on the hyperparameters
- `constr` – sum-to-zero constraint?
- `cyclic` – are you cyclic?
- ...

Example: Add random effect

Add an AR(1) random effect to the linear predictor.

- **Stage 1:**

$$y_i | \eta_i \sim \mathcal{N}(\eta_i, \sigma^2)$$

- **Stage 2:** Covariates and AR(1) component connected to likelihood by

$$\eta_i = \beta_0 + \beta_1 x_i + a_i$$

- **Stage 3:**

- σ^2 : variance of observation noise
- ρ : dependence in AR(1) process
- σ^2 : variance of the innovations in AR(1) process

Example: Add random effect

```
# Generate AR(1) sequence
set.seed(580258)
t = 1:100
rho = 0.8
sd_ar1 = 0.1
ar = rep(0,100)
for(i in 2:100)
  ar[i] = rho * ar[i-1] + rnorm(n = 1, sd = sd_ar1)
# Generate data with AR(1) component
x = runif(100)
y = 1 + 2*x + ar + rnorm(n = 100, sd = 0.2)

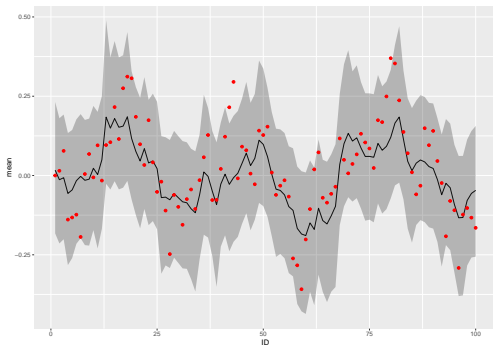
# Run inla
formula = y ~ 1 + x + f(t, model="ar1")

result = inla(formula,
  data = data.frame(x = x, y = y, t = t),
  family = "gaussian")
```

Example

Estimates of the random effect

```
result$summary.random$t %>% ggplot() +  
  geom_line(aes(ID, mean)) +  
  geom_ribbon(aes(ID, ymin = `0.025quant`, ymax = `0.975quant`),  
  geom_point(data = data.frame(t = t , ar = ar), aes(t,ar), color = "red"))
```



Example

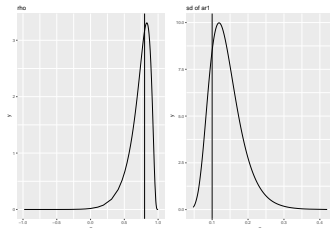
Estimates of the hyperparameters

rho

```
p1 = ggplot() + geom_line(data = data.frame(result$marginals.hyperpar$`
                                aes(x,y)) +
  geom_vline(xintercept = rho) + ggtitle("rho")
```

sd of the ar1 effect

```
prec = result$marginals.hyperpar$`Precision for t`
sd = inla.tmarginal(function(x) 1/sqrt(x), prec)
p2 = ggplot() + geom_line(data = data.frame(sd), aes(x,y)) +
  geom_vline(xintercept = sd_ar1 ) + ggtitle("sd of ar1")
p1+p2
```



Prediction

The interpretation of NA

R-INLA uses NA differently than other packages

- NA in the **response** means no likelihood contribution, i.e. response is unobserved
- NA in a **fixed effect** means no contribution to the linear predictor, i.e. the covariate is set equal to zero
- NA in a **random effect** $f(\dots)$ means no contribution to the linear predictor

Prediction

The distribution of the linear predictor at an unobserved location can be computed by specifying the value of the covariate x and the desired time index t and set y to NA.

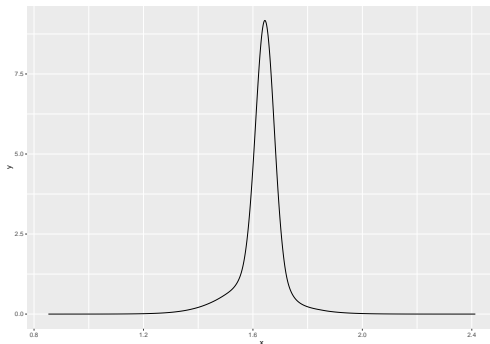
```
# Add one new location
n = 1
x = c(x, runif(n))
t = c(t, 101:(100+n))
y = c(y, rep(NA,n))

# Re-compute
result.pred = inla(formula,
  data = data.frame(x = x, t = t, y = y),
  family="gaussian",
  control.inla = list(int.strategy = "grid"),
  control.compute = list(config = TRUE,
    return.marginals.predictor=TRUE),
  # tell inla to return the marginals for eta!
  control.predictor = list(compute = TRUE))
```

Prediction

Predicted marginal of the linear predictor η_{101}

```
pred = result.pred$marginals.linear.predictor[[100+n]]
pred = inla.smarginal(pred)
ggplot() +
  geom_line(data = data.frame(pred), aes(x, y))
```



Prediction

Caution: This is **not** yet the predictive distribution, as the observation noise is missing.

The predictive distribution is

$$\pi(y_{101}|\mathbf{y})$$

what we got is

$$\pi(\eta_{101}|\mathbf{y})$$

Prediction

One way to add the observation noise to the linear predictor is by sampling from the posterior distribution.

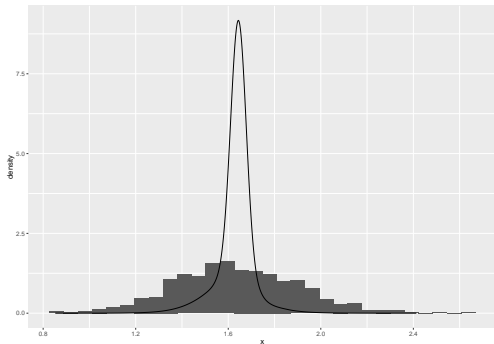
```
n = 1000
x = inla.posterior.sample(n, result.pred)

func = function(...)
{
  eta = Predictor
  eta = eta[101]
  sd = 1/sqrt(theta[1])
  out = rnorm(1, mean = eta, sd =sd)
  return(out)
}

samples = inla.posterior.sample.eval(func, x)[1,]
```

Prediction

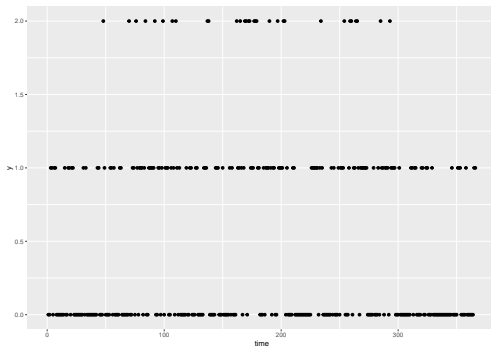
Comparing $\pi(y_{101}|\mathbf{y})$ and $\pi(\eta_{101}|\mathbf{y})$



Smoothing binary time series

Example: Smoothing binary time series

The data set **Tokyo** is available in the **INLA** package and consists of the number of days in Tokyo with rainfall above 1 mm in 1983–1984.



Observations

Each observation consists of

- t : Day of year; $t \in \{1, 2, \dots, 366\}$
- n_t : Number of observations for day t in 1983–1984;
 $n_t \in \{1, 2\}$
- y_t : Number of days with rain out of n_t days for day t ;
 $y_t \in \{0, 1, 2\}$

```
data(Tokyo)
head(Tokyo, 3)
```

```
##    y n time
## 1 0 2    1
## 2 0 2    2
## 3 1 2    3
```

```
Tokyo[60,]
```

```
##    y n time
## 60 0 1   60
```

Hierarchical model

- **Stage 1:** We have binomial responses with known n_t , but unknown probabilities

$$y_t \sim \text{Binomial}(n_t, p_t)$$

- **Stage 2:** A cyclic second order random walk (CRW2) is connected to the likelihood by

$$p_t = \frac{\exp(\eta_t)}{1 + \exp(\eta_t)} \text{ with linear predictor } \eta_t = \text{CRW2}_t$$

- **Stage 3:**
 - τ : Scale parameter in CRW2 with prior

$$\pi(\tau) \sim \text{Gamma}(1, 5 \cdot 10^{-5})$$

INLA implementation

```
# Read data
```

```
data(Tokyo)
```

```
# Specify linear predictor
```

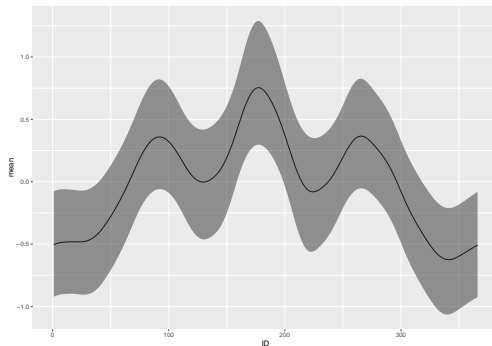
```
formula = y ~ -1 + f(time, model="rw2", cyclic=TRUE)
```

```
# Run model
```

```
result = inla(formula,  
              family = "binomial",  
              Ntrials = n,  
              data = Tokyo)
```

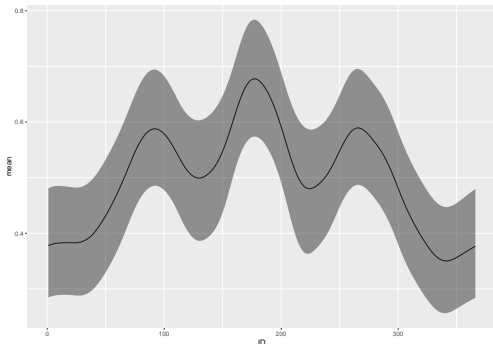
Marginal posterior of CRW2

```
ggplot(data = result$summary.random$t) +  
  geom_line(aes(ID, mean)) +  
  geom_ribbon(aes(ID, ymin = `0.025quant`, ymax = `0.975quant`,  
                    alpha = 0.5))
```



Transform to probability

```
pred = result$summary.fitted.values
pred$ID = 1:dim(Tokyo)[1]
ggplot(data = pred) +
  geom_line(aes(ID, mean)) +
  geom_ribbon(aes(ID, ymin = `0.025quant`, ymax = `0.975quant`),
            alpha = 0.5)
```

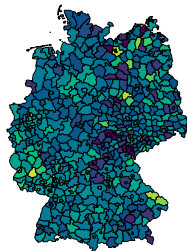


Disease Mapping

Example: disease mapping

We observed larynx cancer mortality counts for males in 544 district of Germany from 1986 to 1990 and want to make a model.

- y_i : The count at location i .
- E_i : An offset; expected number of cases in district i .
- c_i : A covariate (level of smoking consumption) at i
- s_i : spatial location i .



Disease mapping

Assume

$$Y_i \mid \eta_i \sim \text{Poisson}(E_i \exp(\eta_i))$$

where the log relative risk is decomposed into

$$\eta_i = \mu + u_i + v_i$$

- μ is the overall level (intercept).
- $v_i \sim \mathcal{N}(0, \tau_v^{-1})$ represents non-spatial overdispersion.
- u_i are random effects with spatial structure.

A spatially structured effect

To incorporate a spatial structure into a model, the so called **Besag model** is often used.

$$\begin{aligned} p(\mathbf{u} \mid \kappa_{\mathbf{u}}) &\propto \kappa_u^{(n-1)/2} \exp \left(-\frac{\kappa_u}{2} \sum_{i \sim j} (u_i - u_j)^2 \right) \\ &= \kappa_u^{(n-1)/2} \exp \left(-\frac{\kappa_u}{2} \mathbf{u}^T \mathbf{R} \mathbf{u} \right). \end{aligned}$$

where R is called structure matrix and defined as

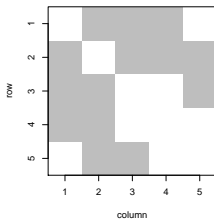
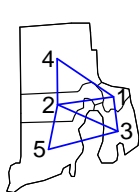
$$R_{ij} = \begin{cases} n_i & i = j \\ -1 & i \sim j \\ 0 & \text{otherwise.} \end{cases}$$

Here, $i \sim j$ denotes that i and j are neighbouring regions.

What does this mean?

Example: Five counties of the US state Rhode Island

The structure matrix \mathbf{R} defines the neighborhood structure.



3	-1	-1	-1	0
-1	4	-1	-1	-1
-1	-1	3	0	-1
-1	-1	0	2	0
0	-1	-1	0	2

Table 1: Structure matrix \mathbf{R}

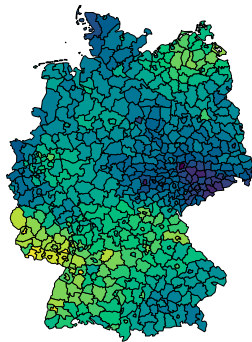
Figure 1: Adjacency matrix

With increasing number of regions \mathbf{R} will be sparse, which allows to do many computations very efficient.

INLA code

```
library(spam)
# load the dataset
data(Oral)
# load the file including neighbourhood information
g = system.file("demodata/germany.graph", package="INLA")
# add one column
Oral = cbind(Oral, region = 1:544, region.unstruc= 1:544)
# define formula
formula = Y ~ f(region, model="besag", graph=g) +
           f(region.unstruc, model="iid")
# run the model
result = inla(formula, family="poisson", E=E, data=Oral)
```

Median of u on exp-scale



Other choices for f-terms

##	[1]	"linear"	"iid"	"mec"	"meb"
##	[6]	"cgeneric"	"rw1"	"rw2"	"crw2"
##	[11]	"besag"	"besag2"	"bym"	"bym2"
##	[16]	"besagproper2"	"fgn"	"fgn2"	"ar1"
##	[21]	"ar"	"ou"	"intslope"	"generic"
##	[26]	"generic1"	"generic2"	"generic3"	"spde"
##	[31]	"spde3"	"iid1d"	"iid2d"	"iid3d"
##	[36]	"iid5d"	"iidkd"	"2diid"	"z"
##	[41]	"rw2diid"	"slm"	"matern2d"	"dmatern"
##	[46]	"clinear"	"sigm"	"revsigm"	"log1exp"

Changing the prior

Changing the prior: Internal scale

- Hyperparameters are represented internally with more well-behaved transformations, e.g. correlation ρ and precision τ are internally

$$\theta_1 = \log(\tau)$$

$$\theta_2 = \log\left(\frac{1+\rho}{1-\rho}\right)$$

- The prior must be set on the parameter in **internal scale**
- Initial values for the mode-search must be set in **internal scale**
- The functions `to.theta()` and `from.theta()` can be used to map back and forth.

Changing the prior: Code

```
hyper = list(prec = list(prior = "loggamma",  
                          param = c(1, 0.1),  
                          initial = 4,  
                          fixed = FALSE))  
  
formula = y ~ f(idx, model = "iid", hyper = hyper) + ...  
  
# For the iid model, default options can be seen with  
inla.doc("iid")
```

Repeated Poisson counts

EPIL example

Seizure counts in a randomised trial of anti-convulsant therapy in epilepsy. From WinBUGS manual.

```
## # A tibble: 6 x 8
##      Ind Repl1 Repl2 Repl3 Repl4   Trt   Base   Age
##    <int> <dbl> <dbl> <dbl> <dbl> <int> <int> <int>
## 1      1      5      3      3      3      0     11     31
## 2      2      3      5      3      3      0     11     30
## 3      3      2      4      0      5      0      6     25
## 4      4      4      4      1      4      0      8     36
## 5      5      7     18      9     21      0     66     22
## 6      6      5      2      8      7      0     27     29
```

Covariates are treatment (0,1), 8-week baseline seizure counts, and age in years.

Repeated Poisson counts

$$\begin{aligned}y_{jk} &\sim \text{Poisson}(\mu_{jk}); \quad j = 1, \dots, 59; \quad k = 1, \dots, 4 \\ \log(\mu_{jk}) &= \alpha_0 + \alpha_1 \log(\text{Base}_j/4) + \alpha_2 \text{Trt}_j \\ &\quad + \alpha_3 \text{Trt}_j \log(\text{Base}_j/4) + \alpha_4 \log(\text{Age}_j) \\ &\quad + \alpha_5 V4 + \text{Ind}_j + \beta_{jk} \\ \alpha_i &\sim \mathcal{N}(0, \tau_\alpha) \quad \tau_\alpha \text{ known (0.001)} \\ \text{Ind}_j &\sim \mathcal{N}(0, \tau_{\text{Ind}}) \quad \tau_{\text{Ind}} \sim \text{Gamma}(1, 0.01) \\ \beta_{jk} &\sim \mathcal{N}(0, \tau_\beta) \quad \tau_\beta \sim \text{Gamma}(1, 0.01)\end{aligned}$$

Here, V4 is an indicator variable for the 4th visit.

Model specification in INLA

The data:

```
##   y Trt Base Age V4 rand Ind
## 1 5   0  11  31  0   1   1
## 2 3   0  11  31  0   2   1
## 3 3   0  11  31  0   3   1
## 4 3   0  11  31  1   4   1
## 5 3   0  11  30  0   5   2
## 6 5   0  11  30  0   6   2
```

The formula:

```
formula = y ~ ClBase4*CTrt + ClAge + CV4 +
  f(Ind, model="iid",
    hyper = list(prec = list(prior = "loggamma",
                             param = c(1,0.01)))) +
  f(rand, model="iid",
    hyper = list(prec = list(prior = "loggamma",
                             param = c(1,0.01))))
```

Run the model:

```
result = inla(formula, family="poisson", data = Epil,
  control.fixed = list(prec.intercept = 0.001,
    prec = 0.001))
```


Comparing results with MCMC

- When comparing the results of R-INLA with MCMC, it is important to use the **same model**.

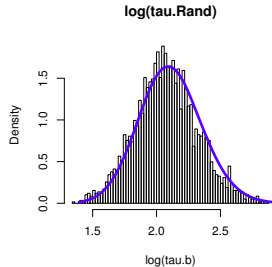
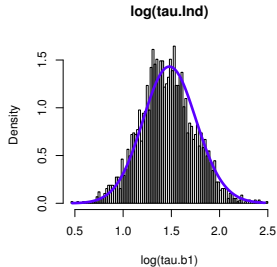
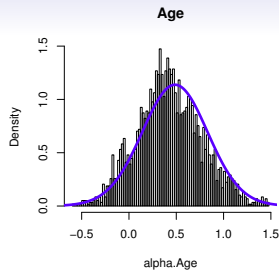
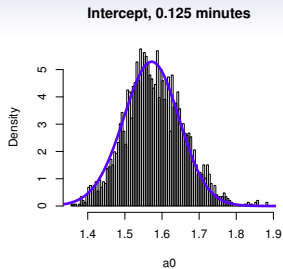
That means, same data, same priors, same constraints on parameters, intercept included or not,

Comparing results with MCMC

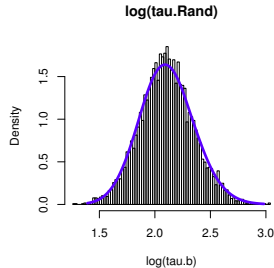
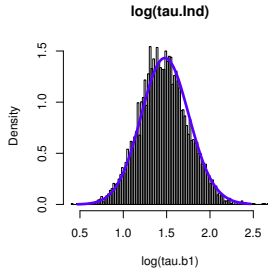
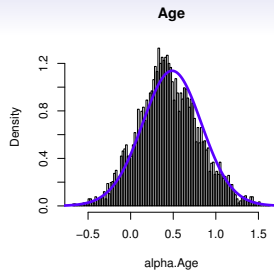
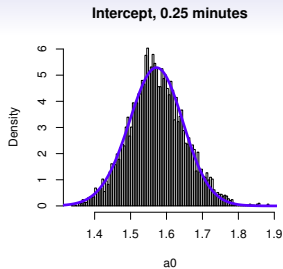
- When comparing the results of R-INLA with MCMC, it is important to use the **same model**.

That means, same data, same priors, same constraints on parameters, intercept included or not,

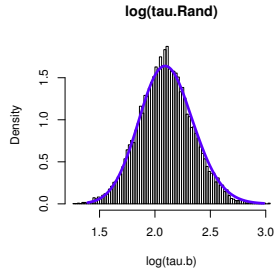
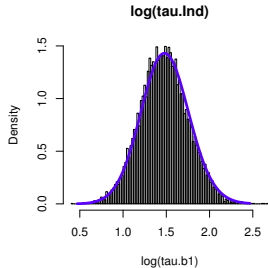
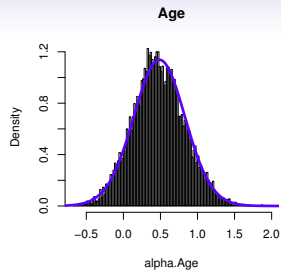
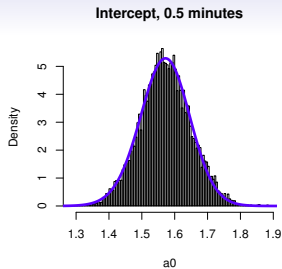
- Here we have compared the results with those obtained using ‘JAGS via the `rjags` package



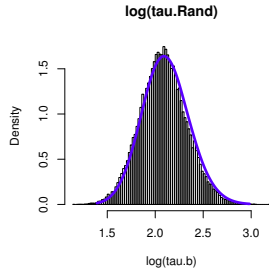
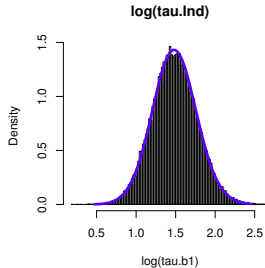
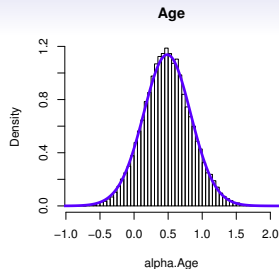
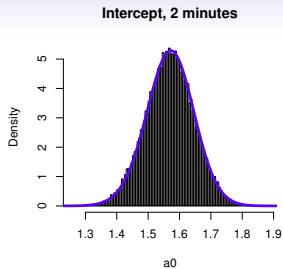
Running time of INLA < 0.5 seconds



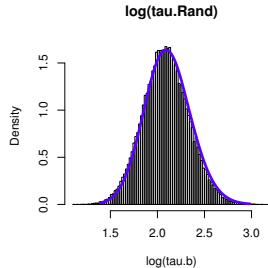
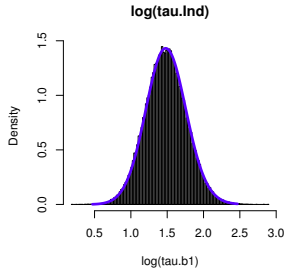
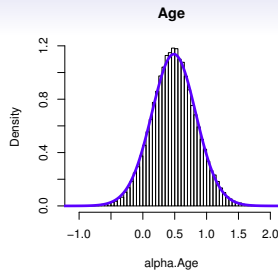
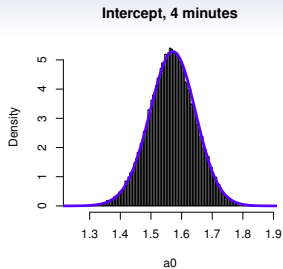
Running time of INLA < 0.5 seconds



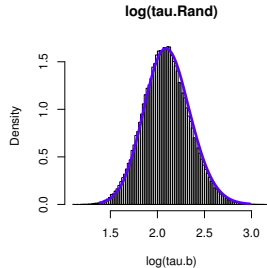
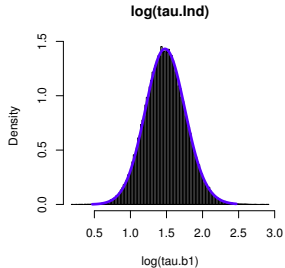
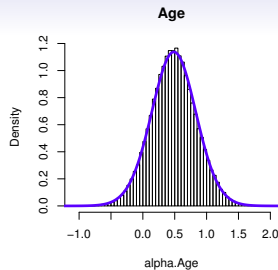
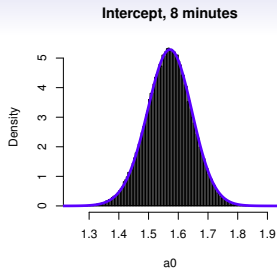
Running time of INLA < 0.5 seconds



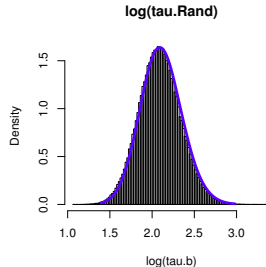
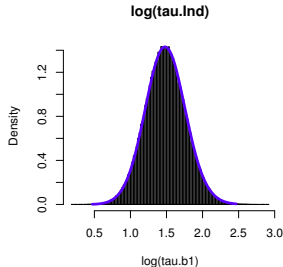
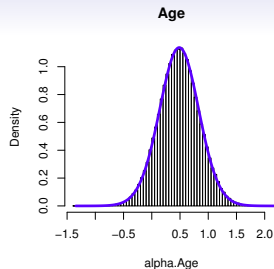
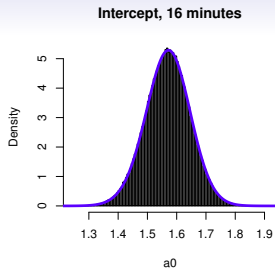
Running time of INLA < 0.5 seconds



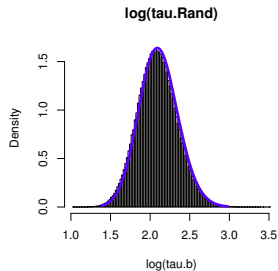
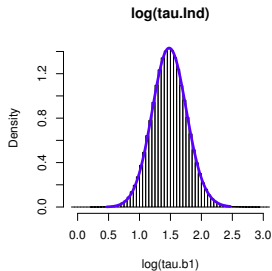
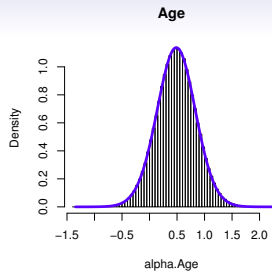
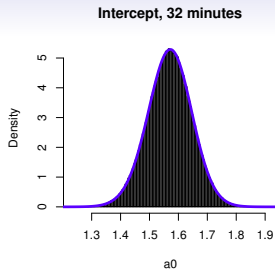
Running time of INLA < 0.5 seconds



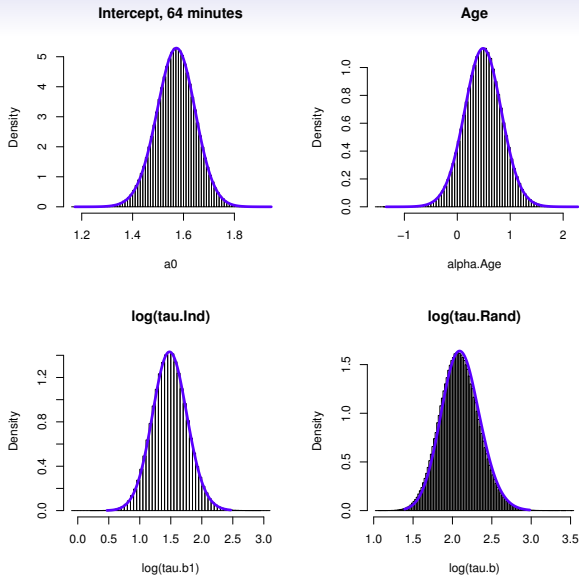
Running time of INLA < 0.5 seconds



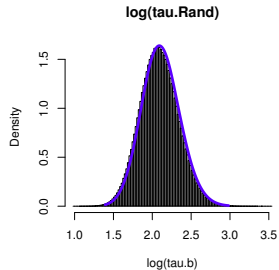
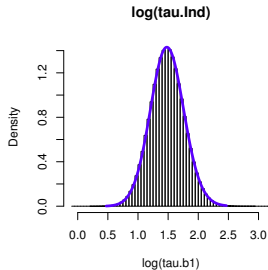
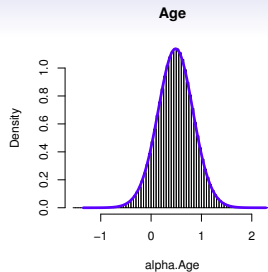
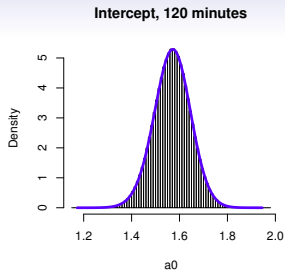
Running time of INLA < 0.5 seconds



Running time of INLA < 0.5 seconds



Running time of INLA < 0.5 seconds



Running time of INLA < 0.5 seconds

Control statements

Control statements

`control.xxx` statements control computations

- `control.fixed`
 - `prec`: Default precision for all fixed effects except the intercept.
 - `prec.intercept`: Precision for intercept (Default: 0.0)
- `control.predictor`
 - `compute`: Compute posterior marginals of linear predictors
- `control.compute`
 - `dic, mlik, cpo`: Compute measures of fit?
 - `config`: Save internal GMRF approximations? (needed to use `inla.posterior.sample()`)
- `control.inla`

`strategy` and `int.strategy` contain useful advanced features
- There are various others as well; see help.

Model Comparison

Model choice/checking

There is a need to compare and choose between various models. This is a difficult problem, but R-INLA has some options available:

- Marginal likelihood \Rightarrow Bayes factors
- Deviance information criterion (DIC)
- Widely applicable information criterion (WAIC)

There are also some predictive checks for the model:

- Conditional predictive ordinate (CPO)
- Probability integral transform (PIT)

Marginal likelihood

```
result = inla(formula,
               data = data,
               control.compute=list(mlik=TRUE))
# See result
result$mlik
```

- Calculates $\log(\pi(\mathbf{y}))$
- Can calculate Bayes factors through differences in value
- **NB:** Problematic for intrinsic models

Deviance information criterion

```
result = inla(formula,
              data = data,
              control.compute=list(dic=TRUE))

# See result
result$dic$dic
```

DIC is a measure of complexity and fit. It is used to compare complex hierarchical models and is defined as:

$$\text{DIC} = \overline{D} + p_D$$

where \overline{D} is the posterior mean of the deviance and p_D is the effective number of parameters. Smaller values of the DIC indicate a better trade-off between complexity and fit of the model.

Widely applicable information criterion (WAIC)

```
result = inla(formula,
              data = data,
              control.compute=list(waic=TRUE))

# See result
result$waic$waic
```

- WAIC is like DIC just newer, and perhaps better
- See “*Understanding predictive information criteria for Bayesian models*” (2013) by Andrew Gelman, Jessica Hwang, and Aki Vehtari

Conditional predictive ordinate

```
result = inla(formula,
              data = data,
              control.compute=list(cpo=TRUE))

# See result
result$cpo$cpo
```

- Measures fit through the predictive density $\pi(y_i^{obs} | \mathbf{y}_{-i})$
- Basically, Bayesian hold-one out
- Easy to compute in the INLA-approach
- Possible failure (`cpofailure`)
- See *Posterior and Cross-validated Predictive Checks: A Comparison of MCMC and INLA* (2009) by Held, Schrödle and Rue

Probability integral transform

```
result = inla(formula,
              data = data,
              control.compute=list(cpo=TRUE))

# See result
result$cpo$pit
```

- Given by

$$\text{Prob}(Y_i \leq y_i^{obs} \mid \mathbf{y}_{-i})$$

- Detects outliers
- Should look out for unusually small or large values
- PIT histograms should be uniform

Model choice: Recap

- Information Criteria or predictive checks essentially for free
- But they may not be appropriate

Thank you for your attention!

If you have any doubts or questions, please write :
sara.martino@math.ntnu.no

