

# N2 - Drug Pressure Plan

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## Day-2 - Friday 19-05-2023

1. Seed Feeders Note: Add 2ml of  $4.8 \times 10^4$  feeder cells / mL cell suspension to each well of 9x 6-well plates

## Day 0 - Saturday 20-05-2023

1. Count N2-Barcode cells and dilute cell suspension to  $5 \times 10^5$  cells/ml
2. Add 1mL of cell suspension/well  
**Note:** Thermofisher indicates that  $1.2 \times 10^6$  cells in a 6 well plate is “confluent” and recommends seeding density of  $0.3 \times 10^6$ . Seeding  $0.5 \times 10^6$  means that after 2 days if dividing normally cells should be close to confluent while adding in extra cells to support those in higher concentrations of drug.
3. Make 1 $\mu$ M working stock of each drug
  - Doxorubicin: 1 $\mu$ L 10mM stock in 10mL media
  - Methotrexate: 1 $\mu$ L 10mM stock in 10mL media
  - Vincristine: 333.33 $\mu$ L 30mM stock in 10mL media

## Plate Maps

**Doxorubicin Dilutions** **Note:** Drug will be diluted by 1/2 when added to well  
Doxorubicin Working Stock (WS) = 1 $\mu$ M

Drug	Drug ID	Dilution volume	Media Volume	Dil 1[Drug]	[Final Drug]
DMSO	DMSO	1 $\mu$ L	9999 $\mu$ L	0	0
Doxorubicin	Dox 1	600 $\mu$ L (from WS)	9700 $\mu$ L	600nm	300nm
Doxorubicin	Dox 2	5000 $\mu$ L (from Dox 1)	5000 $\mu$ L	300nm	150nm
Doxorubicin	Dox 3	5000 $\mu$ L (from Dox 2)	5000 $\mu$ L	150nm	75nm
Doxorubicin	Dox 4	5000 $\mu$ L (from Dox 3)	5000 $\mu$ L	75nm	37.5nm
Doxorubicin	Dox 5	5000 $\mu$ L (from Dox 4)	5000 $\mu$ L	37.5nm	18.75nm

## Doxorubicin Plates

### Dox Plate 1

Well ID	Well Cell Count	Well volume	Drug	Drug Volume	[Final Drug]
A1	5x10 <sup>5</sup>	1 mL	DMSO	1mL	0
A2	5x10 <sup>5</sup>	1 mL	DMSO	1mL	0
A3	5x10 <sup>5</sup>	1 mL	DMSO	1mL	0
B1	5x10 <sup>5</sup>	1 mL	Dox 1	1mL	300nm
B2	5x10 <sup>5</sup>	1 mL	Dox 1	1mL	300nm
B3	5x10 <sup>5</sup>	1 mL	Dox 1	1mL	300nm

### Dox Plate 2

Well ID	Well Cell Count	Well volume	Drug	Drug Volume	[Final Drug]
A1	5x10 <sup>5</sup>	1 mL	Dox 2	1mL	150nm
A2	5x10 <sup>5</sup>	1 mL	Dox 2	1mL	150nm
A3	5x10 <sup>5</sup>	1 mL	Dox 2	1mL	150nm
B1	5x10 <sup>5</sup>	1 mL	Dox 3	1mL	75nm
B2	5x10 <sup>5</sup>	1 mL	Dox 3	1mL	75nm
B3	5x10 <sup>5</sup>	1 mL	Dox 3	1mL	75nm

### Dox Plate 3

Well ID	Well Cell Count	Well volume	Drug	Drug Volume	[Final Drug]
A1	5x10 <sup>5</sup>	1 mL	Dox 4	1mL	37.5nm
A2	5x10 <sup>5</sup>	1 mL	Dox 4	1mL	37.5nm
A3	5x10 <sup>5</sup>	1 mL	Dox 4	1mL	37.5nm
B1	5x10 <sup>5</sup>	1 mL	Dox 5	1mL	18.75nm
B2	5x10 <sup>5</sup>	1 mL	Dox 5	1mL	18.75nm
B3	5x10 <sup>5</sup>	1 mL	Dox 5	1mL	18.75nm

**Methotrexate Dilutions** **Note:** Drug will be diluted by 1/2 when added to well Methotrexate Working Stock = 1 $\mu$ M

Drug	Drug ID	Dilution volume	Media Volume	Dil 1 [Drug]	[Final Drug]
DMSO	DMSO	1 $\mu$ L	9999 $\mu$ L	0	0
Methotrexate	Meth 1	200 $\mu$ L (from WS)	9800 $\mu$ L	200nm	100nm
Methotrexate	Meth 2	5000 $\mu$ L (from Meth 1)	5000 $\mu$ L	100nm	50nm
Methotrexate	Meth 3	5000 $\mu$ L (from Meth 2)	5000 $\mu$ L	40nm	20nm
Methotrexate	Meth 4	5000 $\mu$ L (from Meth 3)	5000 $\mu$ L	20nm	10nm
Methotrexate	Meth 5	5000 $\mu$ L (from Meth 4)	5000 $\mu$ L	10nm	5nm

## Methotrexate Plates

### Meth Plate 1

Well ID	Well Cell Count	Well volume	Drug	Drug Volume	[Final Drug]
A1	5x10 <sup>5</sup>	1 mL	DMSO	1mL	0
A2	5x10 <sup>5</sup>	1 mL	DMSO	1mL	0
A3	5x10 <sup>5</sup>	1 mL	DMSO	1mL	0
B1	5x10 <sup>5</sup>	1 mL	Meth 1	1mL	100nm
B2	5x10 <sup>5</sup>	1 mL	Meth 1	1mL	100nm
B3	5x10 <sup>5</sup>	1 mL	Meth 1	1mL	100nm

### Meth Plate 2

Well ID	Well Cell Count	Well volume	Drug	Drug Volume	[Final Drug]
A1	5x10 <sup>5</sup>	1 mL	Meth 2	1mL	50nm
A2	5x10 <sup>5</sup>	1 mL	Meth 2	1mL	50nm
A3	5x10 <sup>5</sup>	1 mL	Meth 2	1mL	50nm
B1	5x10 <sup>5</sup>	1 mL	Meth 3	1mL	20nm
B2	5x10 <sup>5</sup>	1 mL	Meth 3	1mL	20nm
B3	5x10 <sup>5</sup>	1 mL	Meth 3	1mL	20nm

### Meth Plate 3

Well ID	Well Cell Count	Well volume	Drug	Drug Volume	[Final Drug]
A1	5x10 <sup>5</sup>	1 mL	Meth 4	1mL	10nm
A2	5x10 <sup>5</sup>	1 mL	Meth 4	1mL	10nm
A3	5x10 <sup>5</sup>	1 mL	Meth 4	1mL	10nm
B1	5x10 <sup>5</sup>	1 mL	Meth 5	1mL	5nm
B2	5x10 <sup>5</sup>	1 mL	Meth 5	1mL	5nm
B3	5x10 <sup>5</sup>	1 mL	Meth 5	1mL	5nm

**Vincristine Dilutions** **Note:** Drug will be diluted by 1/2 when added to well  
Vincristine Working Stock = 1 $\mu$ M

Drug	Drug ID	Dilution Volume	Media Volume	Dil 1[Drug]	[Final Drug]
DMSO	DMSO	1 $\mu$ L	9999 $\mu$ L	0	0
Vincristine	Vin 1	8 $\mu$ L (from WS)	9992 $\mu$ L	8nm	4nm
Vincristine	Vin 2	5000 $\mu$ L (from Vin 1)	5000 $\mu$ L	4nm	2nm
Vincristine	Vin 3	5000 $\mu$ L (from Vin 2)	5000 $\mu$ L	2nm	1nm
Vincristine	Vin 4	5000 $\mu$ L (from Vin 3)	5000 $\mu$ L	1nm	0.5nm
Vincristine	Vin 5	5000 $\mu$ L (from Vin 4)	5000 $\mu$ L	0.5nm	0.25nm

## Vincristine Plates

### Vin Plate 1

Well ID	Well Cell Count	Well volume	Drug	Drug Volume	[Final Drug]
A1	5x10 <sup>5</sup>	1 mL	DMSO	1mL	0
A2	5x10 <sup>5</sup>	1 mL	DMSO	1mL	0
A3	5x10 <sup>5</sup>	1 mL	DMSO	1mL	0
B1	5x10 <sup>5</sup>	1 mL	Vin 1	1mL	4nm
B2	5x10 <sup>5</sup>	1 mL	Vin 1	1mL	4nm
B3	5x10 <sup>5</sup>	1 mL	Vin 1	1mL	4nm

### Vin Plate 2

Well ID	Well Cell Count	Well volume	Drug	Drug Volume	[Final Drug]
A1	5x10 <sup>5</sup>	1 mL	Vin 2	1mL	2nm
A2	5x10 <sup>5</sup>	1 mL	Vin 2	1mL	2nm
A3	5x10 <sup>5</sup>	1 mL	Vin 2	1mL	2nm
B1	5x10 <sup>5</sup>	1 mL	Vin 3	1mL	1nm
B2	5x10 <sup>5</sup>	1 mL	Vin 3	1mL	1nm
B3	5x10 <sup>5</sup>	1 mL	Vin 3	1mL	1nm

### Vin Plate 3

Well ID	Well Cell Count	Well volume	Drug	Drug Volume	[Final Drug]
A1	5x10 <sup>5</sup>	1 mL	Vin 4	1mL	0.5nm
A2	5x10 <sup>5</sup>	1 mL	Vin 4	1mL	0.5nm
A3	5x10 <sup>5</sup>	1 mL	Vin 4	1mL	0.5nm
B1	5x10 <sup>5</sup>	1 mL	Vin 5	1mL	0.25nm
B2	5x10 <sup>5</sup>	1 mL	Vin 5	1mL	0.25nm
B3	5x10 <sup>5</sup>	1 mL	Vin 5	1mL	0.25nm

## Day 2 - Monday 22-05-2023

1. Visually check cells for viability
2. Count wells which are growing and split back if overconfluent (>1.0x10<sup>6</sup> cells)
3. Replenish drug media

- Collect and spin down supernatant  
**Note** Add associated drug media to wells while supernatant is being spun down as cells are still attached to feeders
- Resuspend in associated drug media **Note** Repeat drug dilutions from Day 0 except do not make Dil 1 2x [final]

## Day 3 - Tuesday 23-05-2023

1. Seed 9x 6-well feeders
2. Seeding density: Add 2ml of  $4.8 \times 10^4$  feeder cells / mL cell suspension to each well

## Day 4 - Wednesday 24-05-2023

### Collection Day 1

1. Homogenize each well thoroughly
2. Take 1/2 of cell suspension for RNA extraction
  - a. Spin down for 5 min at 250 x g
  - b. Snap freeze in liquid nitrogen
3. Reseed remaining 1/2 of cell suspension
  - a. Count cell suspension
  - b. Reseed  $0.5 \times 10^6$  cells/well (or remaining amount if less than 500K)

## Day 6 - Friday 26-05-2023

1. Visually check cells for viability
  2. Count wells which are growing and split back if overconfluent ( $>1.0 \times 10^6$  cells)
  3. Replenish drug media
- Collect and spin down supernatant  
**Note:** Add associated drug media to wells while supernatant is being spun down as cells are still attached to feeders
  - Resuspend in associated drug media  
**Note** Repeat drug dilutions from Day 0 except do not make Dil 1 2x [final]

## Day 7 - Saturday 27-05-2023

Note: This can be done Friday 1. Seed 9x 6-well feeders 2. Seeding density: Add 2ml of  $4.8 \times 10^4$  feeder cells / mL cell suspension to each well

## Day 8 - Sunday 28-05-2023

### Collection Day 1

1. Homogenize each well thoroughly
2. Take 1/2 of cell suspension for RNA extraction
  - a. Spin down for 5 min at 250 x g

- b. Snap freeze in liquid nitrogen
3. Reseed remaining 1/2 of cell suspension
  - a. Count cell suspension
  - b. Reseed  $0.5 \times 10^6$  cells/well (or remaining amount if less than 500K)

## Day 10 - Tuesday 30-05-2023

1. Visually check cells for viability
2. Count wells which are growing and split back if overconfluent ( $>1.0 \times 10^6$  cells)
3. Replenish drug media
  - Collect and spin down supernatant  
**Note:** Add associated drug media to wells while supernatant is being spun down as cells are still attached to feeders
  - Resuspend in associated drug media  
**Note:** Repeat drug dilutions from Day 0 except do not make Dil 1 2x [final]

## Day 12 - Thursday 1-06-2023

1. Homogenize each well thoroughly
2. Collect entire cell suspension for RNA extraction
  - a. Spin down for 5 min at 250 x g
  - b. Snap freeze in liquid nitrogen