

# Calculating Biological Module Enrichment or Depletion and Visualizing Data on Large-scale Molecular Maps with ACSNMineR and RNavicell Packages

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**Abstract** Biological pathways or modules represent sets of interactions or functional relationships occurring at the molecular level in living cells. A large body of knowledge on pathways is organized in public databases such as the KEGG, Reactome, or in more specialized repositories, the Atlas of Cancer Signaling Network (ACSN) being an example. All these open biological databases facilitate analyses, improving our understanding of cellular systems. We hereby describe **ACSNMineR** for calculation of enrichment or depletion of lists of genes of interest in biological pathways. **ACSNMineR** integrates ACSN molecular pathways gene sets, but can use any gene set encoded as a GMT file, for instance sets of genes available in the Molecular Signatures Database (MSigDB). We also present **RNavicell**, that can be used in conjunction with **ACSNMineR** to visualize different data types on web-based, interactive ACSN maps. We illustrate the functionalities of the two packages with biological data taken from large-scale cancer datasets.

## Introduction

Biological pathways and networks comprise sets of interactions or functional relationships, occurring at the molecular level in living cells (Adriaens et al., 2008; Barillot et al., 2012). A large body of knowledge on cellular biochemistry is organized in publicly available repositories such as the KEGG database (Kanehisa et al., 2011), Reactome (Croft et al., 2014) and MINT (Zanzoni et al., 2002). All these biological databases facilitate a large spectrum of analyses, improving our understanding of cellular systems. For instance, it is a very common practice to cross the output of high-throughput experiments, such as mRNA or protein expression levels, with curated biological pathways in order to visualize the changes, analyze their impact on a network and formulate new hypotheses about biological processes. Many biologists and computational biologists establish list of genes of interest (e.g. a list of genes that are differentially expressed between two conditions, such as normal vs disease) and then evaluate if known biological pathways have significant overlap with this list of genes.

We have recently released the Atlas of Cancer Signaling Network (ACSN), a web-based database which describes signaling and regulatory molecular processes that occur in a healthy mammalian cell but that are frequently deregulated during cancerogenesis (Kuperstein et al., 2015). The ACSN atlas aims to be a comprehensive description of cancer-related mechanisms retrieved from the most recent literature. The web interface for ACSN is using the NaviCell technology, a software framework dedicated to web-based visualization and navigation for biological pathway maps (Kuperstein et al., 2013). This environment is providing an easy navigation of maps through the use of the Google Maps JavaScript library, a community interface with a web blog system, and a comprehensive module for visualization and analysis of high-throughput data (Bonnet et al., 2015).

In this article, we describe two packages related to ACSN analysis and data visualization. The package **ACSNMineR** is designed for the calculation of gene enrichment and depletion in ACSN maps (or any user-defined gene set via the import function), while **RNavicell** is dedicated to data visualization on ACSN maps. Both packages are available on the Comprehensive R Archive Network (<https://cran.r-project.org/web/packages/ACSNMineR/> and <https://cran.r-project.org/web/packages/RNavicell/>), and on the GitHub repository (<https://github.com/sysbio-curie/ACSNMineR> and <https://github.com/sysbio-curie/RNavicell>). For the remainder of this article, we describe the organization of each package and illustrate their capacities with several concrete examples demonstrating their capabilities.

## Packages organization

### ACSNMineR

Currently, ACSN maps cover signaling pathways involved in DNA repair, cell cycle, cell survival, cell death, epithelial-to-mesenchymal transition (EMT) and cell motility. Each of these large-scale

molecular maps is decomposed in a number of functional modules. The maps themselves are merged into a global ACSN map. Thus the information included in ACSN is organized in three hierarchical levels: a global map, five individual maps, and several functional modules. Each ACSN map covers hundreds of molecular players, biochemical reactions and causal relationships between the molecular players and cellular phenotypes. ACSN represents a large-scale biochemical reaction network of 4,826 reactions involving 2,371 proteins (as of today), and is continuously updated and expanded. We have included the three hierarchical levels in the **ACSNMineR** package, in order to be able to calculate enrichments at all three levels. The calculations are made by counting the number of occurrences of gene symbols (HUGO gene names) from a given list of genes of interest in all ACSN maps and modules. Table 1 is detailing the number of gene symbols contained in all the ACSN maps.

**Table 1:** ACSN maps included in the **ACSNMineR** package. Map: map name, Total: total number of gene symbols (HUGO) used to construct the map, Nb mod.: number of modules, Min: minimum number of gene symbols in the modules, Max: maximum number of gene symbols in the modules, Mean: average number of gene symbols per module. N.B.: one gene symbol may be present in several modules of the map.

Map	Total	Nb mod.	Min	Max	Mean
ACSN global	2239	67	2	629	79
Survival	1053	5	208	431	328
Apoptosis	667	7	19	382	136
EMT & Cell motility	634	9	18	629	137
DNA repair	345	21	3	171	45
Cell cycle	250	25	2	130	20

The statistical significance of the counts in the modules is assessed by using either the Fisher exact test (Fisher, 1922, 1934) or the hypergeometric test, which are equivalent for this purpose (Rivals et al., 2007).

The current ACSN maps are included in the **ACSNMineR** package, as a list of character matrices.

```
> length(ACSN_maps)
[1] 6
> names(ACSN_maps)
[1] "Apoptosis"    "CellCycle"    "DNA_repair"   "EMT_motility" "Master"
[6] "Survival"
```

For each matrix, rows represent a module, with the name of the module in the first column, followed by a description of the module (optional), and then followed by all the gene symbols of the module. The maps will be updated according to every ACSN major release.

The main function of the **ACSNMineR** package is the enrichment function, which is calculating over-representation or depletion of genes in the ACSN maps and modules. We have included a small list of 12 Cell Cycle related genes in the package, named `genes_test` that can be used to test the main enrichment function and to get familiar with its different options.

```
> genes_test
[1] "ATM"    "ATR"    "CHEK2"  "CREBBP" "TFDP1"  "E2F1"   "EP300"
[8] "HDAC1"  "KAT2B"  "GTF2H1" "GTF2H2" "GTF2H2B"
```

The example shown below is the simplest command that can be done to test a gene list for over-representation on the six included ACSN maps. With the list of 12 genes mentioned above and a default p-value cutoff of 0.05, we have a set of 8 maps or modules that are significantly enriched. The results are structured as a data frame with nine columns displaying the module name, the module size, the number of genes from the list in the module, the names of the genes that are present in the module, the size of the reference universe, the number of genes from the list that are present in the universe, the raw p-value, the p-value corrected for multiple testing and the type of test performed. The module field in the results data frame indicate the map name and the module name separated by a column character. If a complete map is significantly enriched or depleted, then only the map name is shown, without any module or column character. For instance, the third line of the results object below concern the E2F1 module of the CellCycle map.

```
> library(ACSNMineR)
> results <- enrichment(genes_test)
> dim(results)
```

```
[1] 8 9
> results[3,]
      module module_size nb_genes_in_module
V161 CellCycle:E2F1      19                12
                                genes_in_module
V161 ATM ATR CHEK2 CREBBP TFDP1 E2F1 EP300 HDAC1 KAT2B GTF2H1 GTF2H2 GTF2H2B
      universe_size nb_genes_in_universe      p.value p.value.corrected      test
V161             2237                   12 3.735018e-21    2.353061e-19 greater
```

The enrichment function can take up to nine arguments: the gene list (as a character vector), the list of maps that will be used to calculate enrichment or depletion, the type of statistical test (either the Fisher exact test or the hypergeometric test), the module minimal size for which the calculations will be done, the universe, the p-value threshold, the alternative hypothesis ("greater" for calculating over-representation, "less" for depletion and "both" for both tests) and a list of genes that should be removed from the universe (option "Remove\_from\_universe"). This option may be useful for instance if we know beforehand that a number of genes are not expressed in the samples considered.

Only the gene list is mandatory to call the enrichment function, all the other arguments have default values. The maps argument can either be a dataframe imported from a GMT file with the `format_from_gmt` function or a list of dataframes generated by the same procedure. The GMT format corresponds to the Broad Institute's Gene Matrix Transposed file format, a convenient and easy way to encode named sets of genes of interest in tab-delimited text files (it is not a graph or network format). By default, the function enrichment uses the ACSN maps previously described. The correction for multiple testing is set by default to use the method of Benjamini & Hochberg, but can be changed to any of the usual correction methods (Bonferroni, Holm, Hochberg, Holm, or Benjamini & Yekutieli (Reiner et al., 2003)), or even disabled. The minimal module size represents the smallest size value of a module that will be used to compute enrichment or depletion. This is meant to remove results of low significance for module of small size. The universe in which the computation is made by default is defined by all the gene symbols contained in the maps. All the genes that were given as input and that are not present on the maps will be discarded. To keep all genes, the user can change the universe to HUGO, and in that case, the complete list of HUGO gene symbols will be used as the reference (> 39,000 genes). The threshold corresponds to the maximal value of the corrected p-value (unless the user chose not to correct for multiple testing) that will be displayed in the result table.

It may be of interest to compare enrichment of pathways in different cohorts or experiments. For example, enrichment of highly expressed pathways can reveal differences between two cancer types or two cell lines. To facilitate such comparisons, **ACSNMineR** provides a `multisample_enrichment` function. It relies on the enrichment function but takes a list of character vector genes. The name of each element of the list will be assumed to be the name of the sample for further analysis. Most of the arguments given to `multisample_enrichment` are the same as the ones passed to `enrichment`. However, the `cohort_threshold` is designed to filter out modules which would not pass the significance threshold in all samples.

Finally, to facilitate visualization of results, **ACSNMineR** integrates a representation function based on `ggplot2` syntax (Wickham, 2009). It allows representation of results from enrichment or `multisample_enrichment` with a limited number of parameters. Two types of display are available: heat-map tiles or bars. For multiple samples using a barplot representation, the number of rows used can be provided, otherwise all plots will be on the same row. For the heatmap, the color of the non-significant modules, and boundaries of the gradient for significant values can also be tuned.

We previously computed the p-value of the `genes_test` list with default parameters. The number of modules which have a p-value below 0.05 was 8, that can be compared to the 16 obtained without correction with the simple command shown below (some of the results are displayed in table 2).

```
enrichment(genes_test, correction_multitest = FALSE)
```

We can now plot the first six rows of the results obtained for corrected and uncorrected fisher test with heatmap format (Figure 1) or barplot (Figure 2) with the following commands:

```
# heatmap

represent_enrichment(enrichment = list(Corrected = results[1:6,],
Uncorrected = results_uncorrected[1:6,]),
                      plot = "heatmap", scale = "reverselog",
                      low = "steelblue", high = "white", na.value = "grey")

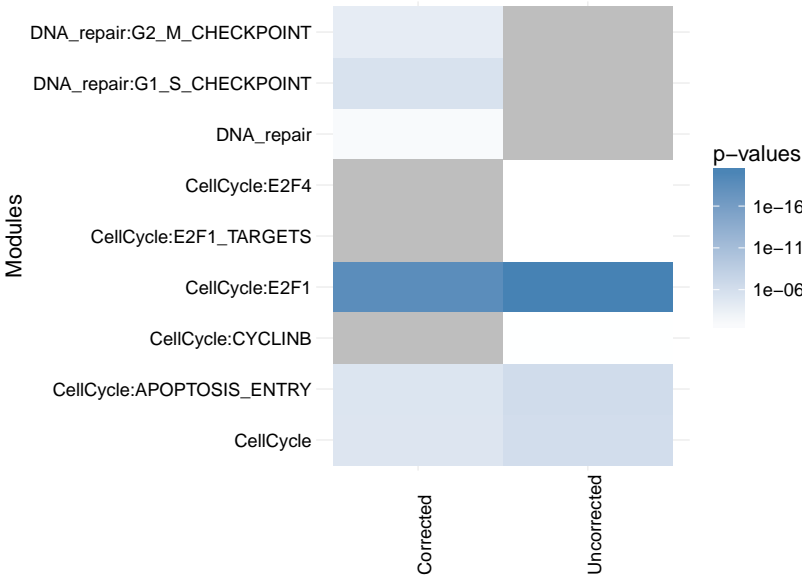
# barplot
```

**Table 2:** First rows of the results from enrichment analysis without correction. Module : name of the module. Mod. size: size of the module. Genes in module: genes from input which are found in the module. p-value: uncorrected p-value. Test : null hypothesis used, greater is synonym of enrichment.

Module	Mod. size	Genes in module	p-value	Test
CellCycle	242	ATM ATR CHEK2 CREBBP TFDP1 E2F1 EP300 HDAC1 KAT2B GTF2H1 GTF2H2 GTF2H2B	$5.4 \times 10^{-7}$	greater
CellCycle:APOPTOSIS_ENTRY	10	ATM ATR CHEK2 E2F1	$3.5 \times 10^{-7}$	greater
CellCycle:CYCLINB	7	ATM	0.04	greater

```
represent_enrichment(enrichment = list(Corrected = results[1:6,],
                                       Uncorrected = results_uncorrected[1:6,]),
                    plot = "bar", scale = "reverselog",
                    nrow = 1)
```

**Figure 1:** Representation of the enriched modules (first six rows for each setting), with either Bonferroni correction or no correction. Grey tiles means that the data is not available for this module in this sample. P-values of low significance are in white, whereas p-values of high significance are represented in blue.

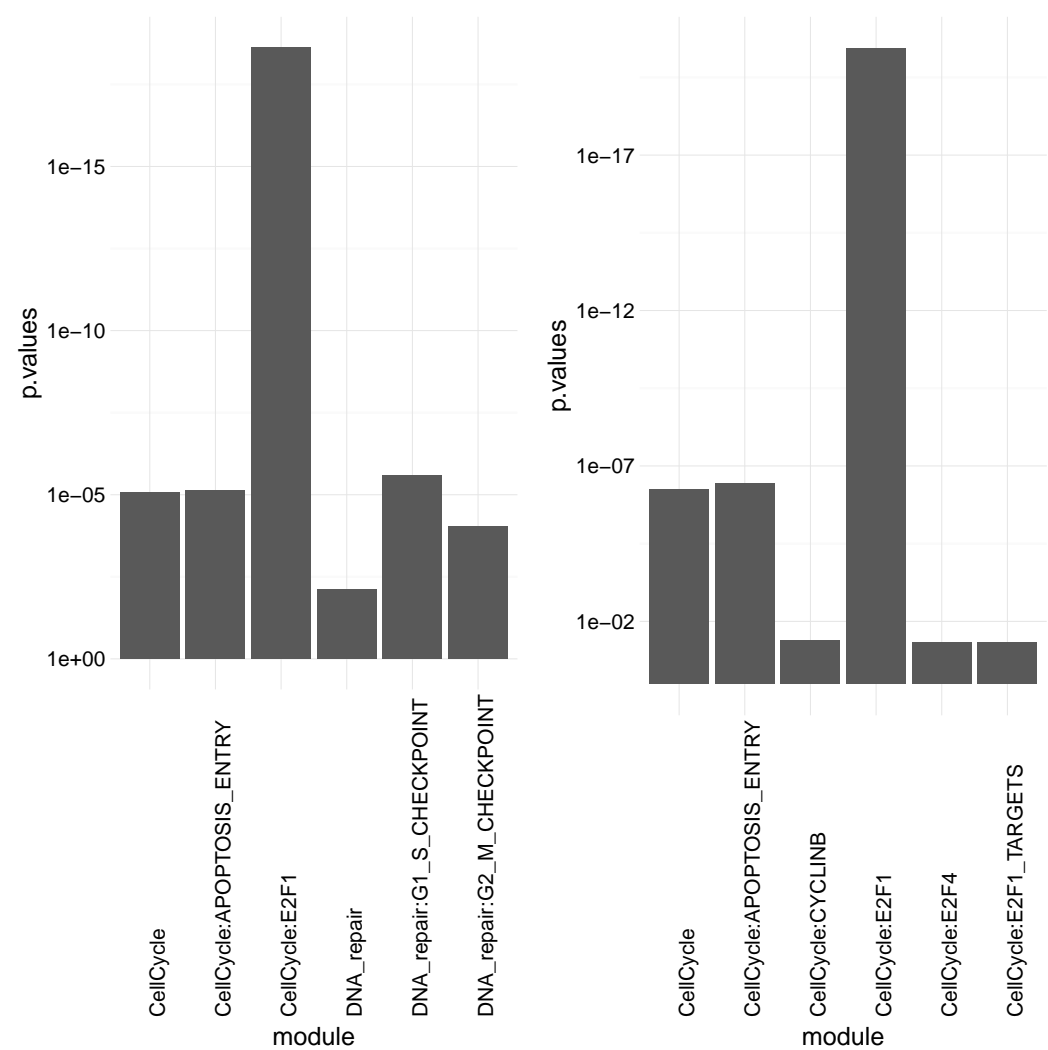


RNaviCell

The NaviCell Web Service provides a server mode, which allows automating visualization tasks and retrieving data from molecular maps via RESTful (standard http/https) calls. Bindings to different programming languages are provided in order to facilitate the development of data visualization workflows and third-party applications (Bonnet et al., 2015). RNaviCell is the R binding to the NaviCell Web Service. It is implemented as a standard R package, using the R object-oriented framework known as Reference Classes (Wickham, 2015). Most of the work done by the user using graphical point-and-click operations on the NaviCell web interface are encoded as functions in the library encapsulating http calls to the server with appropriate parameters and data. Calls to the NaviCell server are performed using the library RCurl (Lang and the CRAN team, 2015), while data encoding/decoding in JSON format is performed with the RJSONIO library (Lang, 2014).

Once the RNaviCell library is installed and loaded, the first step is to create a NaviCell object and launch the browser session. This will automatically create a unique session ID with the NaviCell server. Once the session is established, various functions can be called to send data to the web

**Figure 2:** Representation of the enriched modules (first six rows for each setting), with either Bonferroni correction (left) or no correction (right). The modules are on the X axis and the p-values are on the Y axis.



session, set graphical options, visualize data on a map or get data from the map. There are 125 functions available in the current version of RNavicell. All of them are described with their different options in the RNavicell documentation, and we provide a tutorial on the GitHub repository wiki (<https://github.com/sysbio-curie/RNavicell/wiki/Tutorial>).

In the simple example detailed below, we create a NaviCell session, then load an expression data set from a local (tab-delimited) file. The data represent gene expression measured in a prostate cancer cell line resistant to hormonal treatment (agressive), and is taken from the Cell Line Encyclopedia project (Barretina et al., 2012). We visualize the data values on the Cell Cycle map (the default map), using heat maps. With this visualization mode, gene expression values are represented as a color gradient (green to red) in squares positioned next to the entities where the gene has been mapped (Figure 3). Note that the map is displayed in a browser and is *interactive*, i.e. users can zoom in to display more information and for example look in what reactions are involved the genes selected to be displayed, and lots of other informations (see Bonnet et al. (2015) and Kuperstein et al. (2015) for more details).

```
# a short RNavicell script example

# load RNavicell library

library(RNavicell)
```

ISSN 2073-4859



additional prognostic motifs (Schmidt et al., 2008). Estrogen receptor (ER) expression and proliferative activity of breast carcinomas are well-known and described prognostic markers. Patients with ER-positive carcinomas have a better prognosis than those with ER-negative carcinomas, and rapidly proliferating carcinomas have an adverse prognosis. Knowledge about the molecular mechanisms involved in the processes of estrogen-dependent tumor growth and proliferative activity has led to the successful development of therapeutic concepts, such as antiendocrine and cytotoxic chemotherapy.

The dataset corresponding to this study is available as a Bioconductor package. The code shown below is creating a list of differentially expressed genes between ER positive and ER negative samples, and calculates the enrichment in ACSN maps from this list of genes. As seen in Table 3, there is one map (DNA repair) and seven modules (belonging to the Cell Cycle, DNA repair and Apoptosis maps) enriched.

```
# load all necessary packages
library(breastCancerMAINZ)
library(Biobase)
library(limma)
library(ACSNMineR)
library(hgu133a.db)
library(RNaviCell)

# load data and extract expression and phenotype data
data(mainz)
eset <- exprs(mainz)
pdat <- pData(mainz)

# Create list of genes differentially expressed between ER positive and
# ER negative samples using moderated t-test statistics
design <- model.matrix(~factor(pdat$er == '1'))
lmFit(eset, design) -> fit
eBayes(fit) -> ebayes
topTable(ebayes, coef=2, n=25000) -> tt
which(tt$adj < 0.05) -> selection
rownames(tt[selection,]) -> probe_list
mget(probe_list, env = hgu133aSYMBOL) -> symbol_list
symbol_list <- as.character(symbol_list)

# calculate enrichment in ACSN maps

enrichment(symbol_list) -> results

dim(results)
[1] 8 9
```

**Table 3:** ACSN maps enrichment for genes differentially expressed between ER positive and ER negative samples in breast cancer. Module : name of the map/module. Mod. size: size of the module. Nb genes: number of genes from input which are found in the module. pval: raw p-value. Cor. pval: corrected p-value.

Module	Mod. size	Nb genes	pval	Cor. pval
Apoptosis:AKT_MTOR	79	47	0.00043	0.0068
CellCycle:E2F2_TARGETS	35	22	0.0055	0.043
CellCycle:E2F3_TARGETS	51	31	0.0023	0.025
CellCycle:E2F4_TARGETS	100	60	$5.8 \times 10^{-5}$	0.0037
DNA_repair	346	172	0.00038	0.0068
DNA_repair:CELL_CYCLE	82	49	0.00029	0.0068
DNA_repair:G1_CC_PHASE	25	18	0.0013	0.016
DNA_repair:S_CC_PHASE	46	28	0.0036	0.033

The Molecular Signatures Database (MSigDB) is one of the most widely used repository of well-annotated gene sets representing the universe of biological processes (Liberzon et al., 2011). We downloaded the canonical pathways set, counting more than 1,300 gene sets representing canonical pathways compiled by domain experts. The dataset is encoded with the GMT format, and can be

imported within ACSNMiner with the `format_from_gmt` function. We calculate the enrichment for the breast cancer differentially expressed gene list, simply specifying the MSigDB data we just imported as the `maps` option. Table 4 is displaying the pathways having a corrected p-value < 0.05. The prefix is indicating the database source, so we see that we have pathways from the KEGG, Reactome and PID databases. Consistent with our previous results, most of the enriched pathways are related to the cell cycle regulation.

```
# Import MSigDB canonical pathways and calculate enrichment on this database
```

```
mtsig <- format_from_gmt('c2.cp.v5.0.symbols.gmt')
enrichment(symbol_list, maps = mtsig)
```

**Table 4:** MSigDB canonical pathway database enrichment for genes differentially expressed between ER positive and ER negative samples in breast cancer. This table presents the 10 modules with lowest p-value out of 125 with corrected p-value lower than 0.05. Module : name of the module. Mod. size: size of the module. Nb genes: number of genes from input which are found in the module. Cor. pval: corrected p-value.

Pathway	Mod. size	Nb genes	Cor. pval
KEGG_CELL_CYCLE	128	76	$3.9 \times 10^{-8}$
REACTOME_CELL_CYCLE_MITOTIC	325	159	$3.9 \times 10^{-8}$
REACTOME_DNA_REPLICATION	192	98	$4.9 \times 10^{-6}$
PID_FOXM1PATHWAY	40	29	$3.1 \times 10^{-5}$
REACTOME_MITOTIC_M_M_G1_PHASES	172	87	$3.1 \times 10^{-5}$
REACTOME_CELL_CYCLE	421	182	$5 \times 10^{-5}$
REACTOME_MITOTIC_G1_G1_S_PHASES	137	71	$9 \times 10^{-5}$
PID_AURORA_B_PATHWAY	39	27	0.0002
REACTOME_S_PHASE	109	58	0.00024
PID_SYNDECAN_1_PATHWAY	46	30	0.00026

At last, we visualize the mean expression values for ER negative samples for all genes differentially expressed on the ACSN master (global) map using RNavicell commands to create heatmaps.

```
# Select ER negative samples and calculate mean expression values
```

```
apply(eset[probe_list, pdat$er == 0], 1, mean) -> er_minus_mean
names(er_minus_mean) <- symbol_list
er_minus_mean <- as.matrix(er_minus_mean)
colnames(er_minus_mean) <- c('exp')
```

```
# create a NaviCell session, import the expression matrix on the map and create
# heatmaps to represent the data points.
```

```
navicell <- NaviCell()
navicell$proxy_url <- "https://acsn.curie.fr/cgi-bin/nv_proxy.php"
navicell$map_url <- "https://acsn.curie.fr/navicell/maps/acsn/master/index.php"

navicell$launchBrowser()
navicell$importDatatable("mRNA expression data", "GBM_exp", er_minus_mean)
navicell$heatmapEditorSelectSample('0', 'exp')
navicell$heatmapEditorSelectDatatable('0', 'GBM_exp')
navicell$heatmapEditorApply()
```

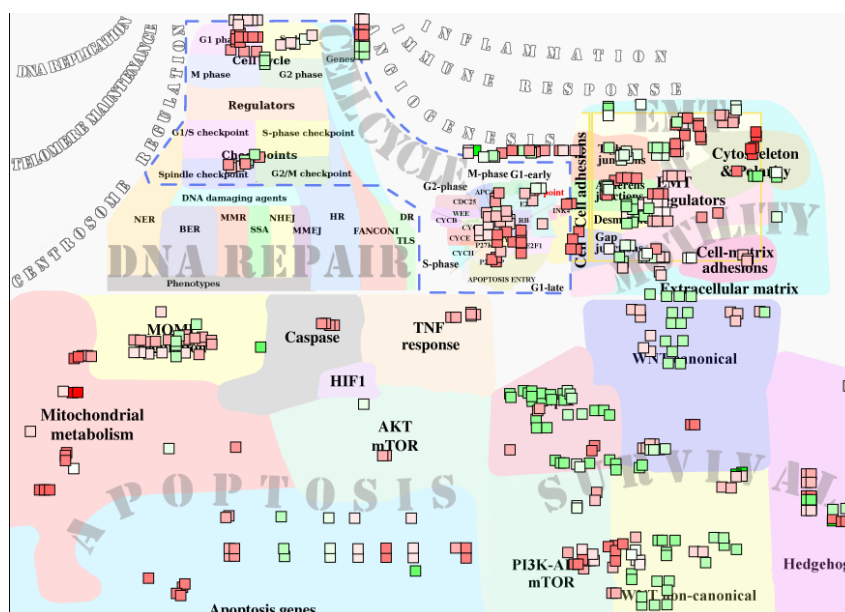
The Figure 4 is displaying the map for genes having a corrected p-value < 0.05.

### Analysis of glioblastoma mutation frequencies

Recent years have witnessed a dramatic increase in new technologies for interrogating the activity levels of various cellular components on a genome-wide scale, including genomic, epigenomic, transcriptomic, and proteomic information (Hawkins et al., 2010). Integrating these heterogeneous datasets



**Figure 4:** Mean expression values for ER negative differentially expressed genes in breast cancer visualized as heatmaps on the ACSN master map.



provides more biological insights than performing separate analyses. For instance, international consortia such as The Cancer Genome Atlas (TCGA) have launched large-scale initiatives to characterize multiple types of cancer at different levels on hundreds of samples. These integrative studies have already led to the identification of novel cancer genes (McLendon et al., 2008).

Malignant gliomas, the most common subtype of primary brain tumors, are aggressive, highly invasive, and neurologically destructive tumors considered to be among the deadliest of human cancers. In its most aggressive manifestation, glioblastoma (GBM), median survival ranges from 9 to 12 months, despite maximum treatment efforts (Maher et al., 2001). In this study we have analyzed whole-genome mutation data generated by the TCGA project on hundreds of patients. More specifically, we parsed the MAF (Mutation Annotation Format) GBM files produced by different sequencing centers to count and calculate gene mutation frequencies. We kept the mutations having a status likely to disturb the target protein's function (i.e. Frame\_Shift\_Del, Nonstop\_Mutation, In\_Frame\_Del, In\_Frame\_Ins, Missense\_Mutation, Nonsense\_Mutation, Splice\_Site, Translation\_Start\_Site). In total, we collected mutations for more than 13,000 genes in a total of 379 mutated samples. In order to retain the most frequently mutated genes, we calculated frequencies across all mutated samples, and kept genes having a frequency greater than 1% (3,293 genes). We further labelled genes having a frequency greater than 1% and less than 5% as "1" and genes highly mutated (frequency higher than 5%) as "2".

We loaded the data as a matrix in R and calculated the enrichment in ACSN maps with the ACSN-MineR function `enrichment`. The results are displayed in table 5. There are 6 modules significantly enriched in the DNA repair and EMT motility maps. Cell matrix adhesions and ECM (extra cellular matrix), part of the EMT motility map, are the modules with highest significance. The EMT motility map is significantly enriched at the global map level (second line in the table).

**Table 5:** ACSN maps enrichment for frequently mutated glioblastoma genes. Module : name of the module. Mod. size: size of the module. Nb genes: number of genes from input which are found in the module. Cor. pval: corrected p-value.

module	Mod. size	Nb genes	Cor. pval
DNA_repair:S_PHASE_CHECKPOINT	45	19	0.008
EMT_motility	635	181	0.0002
EMT_motility:CELL_MATRIX_ADHESIONS	73	45	3.73e-12
EMT_motility:CYTOSKELETON_POLARITY	154	47	0.022
EMT_motility:DESMOSOMES	29	15	0.002
EMT_motility:ECM	147	69	9.77e-11
EMT_motility:EMT_REGULATORS	629	178	0.0002

Visualization of the list of glioblastoma mutated genes is shown on figure 5. This figure was

generated with the ACSNMineR commands detailed below. Results of the enrichment test correlate well with the visualization on the map, with a high density of low and high frequency mutated genes in the EMT motility and DNA repair regions (maps) of the global ACSN map. Although they are not statistically significant, quite high densities can also be seen in other regions of the map.

```
library(RNaviCell)

# Create a NaviCell object, point it to the ACSN master map and launch
# a session.

navicell <- NaviCell()
navicell$proxy_url <- "https://acsn.curie.fr/cgi-bin/nv_proxy.php"
navicell$map_url <- "https://acsn.curie.fr/navicell/maps/acsn/master/index.php"
navicell$launchBrowser()

# Read the GBM data file and import it into the session.

mat <- navicell$readDatatable('gbm.txt')
navicell$importDatatable("Mutation data", "GBM", mat)

# set datatable and sample names for the glyph editor

navicell$drawingConfigSelectGlyph(1, TRUE)
navicell$glyphEditorSelectSample(1, "categ")
navicell$glyphEditorSelectShapeDatatable(1, "GBM")
navicell$glyphEditorSelectColorDatatable(1, "GBM")
navicell$glyphEditorSelectSizeDatatable(1, "GBM")
navicell$glyphEditorApply(1)

# set color, shape and size parameters for glyphs

navicell$unorderedConfigSetDiscreteShape("GBM", "sample", 0, 1)
navicell$unorderedConfigSetDiscreteShape("GBM", "sample", 1, 5)
navicell$unorderedConfigApply("GBM", "shape")

navicell$unorderedConfigSetDiscreteColor("GBM", "sample", 0, "398BC3")
navicell$unorderedConfigSetDiscreteColor("GBM", "sample", 1, "CC5746")
navicell$unorderedConfigApply("GBM", "color")

navicell$unorderedConfigSetDiscreteSize("GBM", "sample", 0, 4)
navicell$unorderedConfigSetDiscreteSize("GBM", "sample", 1, 14)

navicell$unorderedConfigApply("GBM", "size")
```

## Summary and perspectives

In this work, we presented the R package **ACSNMineR**, a novel package for the calculation of p-values for enrichment or depletion of genes in biological pathways. The package includes the six large-scale molecular maps and 67 functional modules of the Atlas of Cancer Signaling Network (ACSN). Enrichment can be calculated for those maps and modules with several options to play with, but can also be calculated for other databases of molecular pathways, that can be imported from GMT formatted files.

We also describe in this work the **RNaviCell** package, a R package convenient to use with **ACSN-MineR**. This package is dedicated to create web-based and interactive data visualization on ACSN maps. Users can use this tools to represent genes of interest that have been shown to be related to the maps by calculating enrichment with **ACSNMineR**. Creating maps with the graphical user interface of the ACSN website can be a tedious task if the user has multiple samples or gene lists, and wants to compare their representations on ACSN maps. The **RNaviCell** package can be used to automate the process of creating the graphical representations automatically. The maps are displayed in a browser and are interactive, with the possibility for the user to zoom in and out, search for genes or molecular species, and see the details of the molecular reactions (what partners are involved, what is the state of a given species, etc.). For more details on how to use the interface and the different possibilities, see [Kuperstein et al. \(2013\)](#), [Bonnet et al. \(2015\)](#) and [Kuperstein et al. \(2015\)](#). We have shown how



## Acknowledgements

This work was supported by a grant “Projet Incitatif et Collaboratif Computational Systems Biology Approach for Cancer” from Institut Curie. The authors would like to thank Pierre Gestraud for his comments on early versions of the **ACSNMineR** package and Eric Viara for guidance and assistance on the development of the **RNaviCell** package.

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