

FMPH221 Project Report Code

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Loading Libraries

```
library(alr4)

## Loading required package: car
## Loading required package: carData
## Loading required package: effects
## Warning: package 'effects' was built under R version 4.1.1
## lattice theme set by effectsTheme()
## See ?effectsTheme for details.

library(GGally)

## Warning: package 'GGally' was built under R version 4.1.1
## Loading required package: ggplot2
## Warning: package 'ggplot2' was built under R version 4.1.1
## Registered S3 method overwritten by 'GGally':
##   method from
##   +.gg      ggplot2

library(leaps)
```

Functions

```
normalize = function(x) { # used to scale data to 0 to 1 [later important for log transformation]
  (x - min(x)) / (max(x) - min(x))
}
```

Data and key

```
neuromaps_df = read.csv("/Users/kevinnguyen/Desktop/fmph221/data/neuromaps-mni152_y-siips_res-basc444.c
neuromaps_df = apply(neuromaps_df, 2, normalize) |> as.data.frame()
neuromaps_key = read.csv("/Users/kevinnguyen/Desktop/fmph221/data/neuromaps-key.csv")
```

Log transforming the skewed variables

Chosen by a visual inspection of each histogram

```
logcols = c(
  "aghourian2017.feobv",
  "bedard2019.feobv",
  "gallezot2010.p943",
  "jaworska2020.fallypride",
  "sandiego2015.flb457",
  "sasaki2012.fepe2i",
  "tuominen.feobv"
)
neuromaps_df[logcols] = log(neuromaps_df[logcols] + 1)
```

Principal Component Analyses to reduce collinearity and dimensionality

Many of the predictors are measuring the same things

```
neuromaps_pca = data.frame(siips = neuromaps_df$siips)

for (group in unique(neuromaps_key$description)) {
  colnames = neuromaps_key[neuromaps_key$description == group, "colname"]
  if (length(colnames) > 1) { # first PC
    pcs <- prcomp(neuromaps_df[, colnames])
    neuromaps_pca[, paste0(group, ".pc1")] = pcs$x[, 1]
  } else { # leave as is
    neuromaps_pca[group] = neuromaps_df[, colnames]
  }
}
```

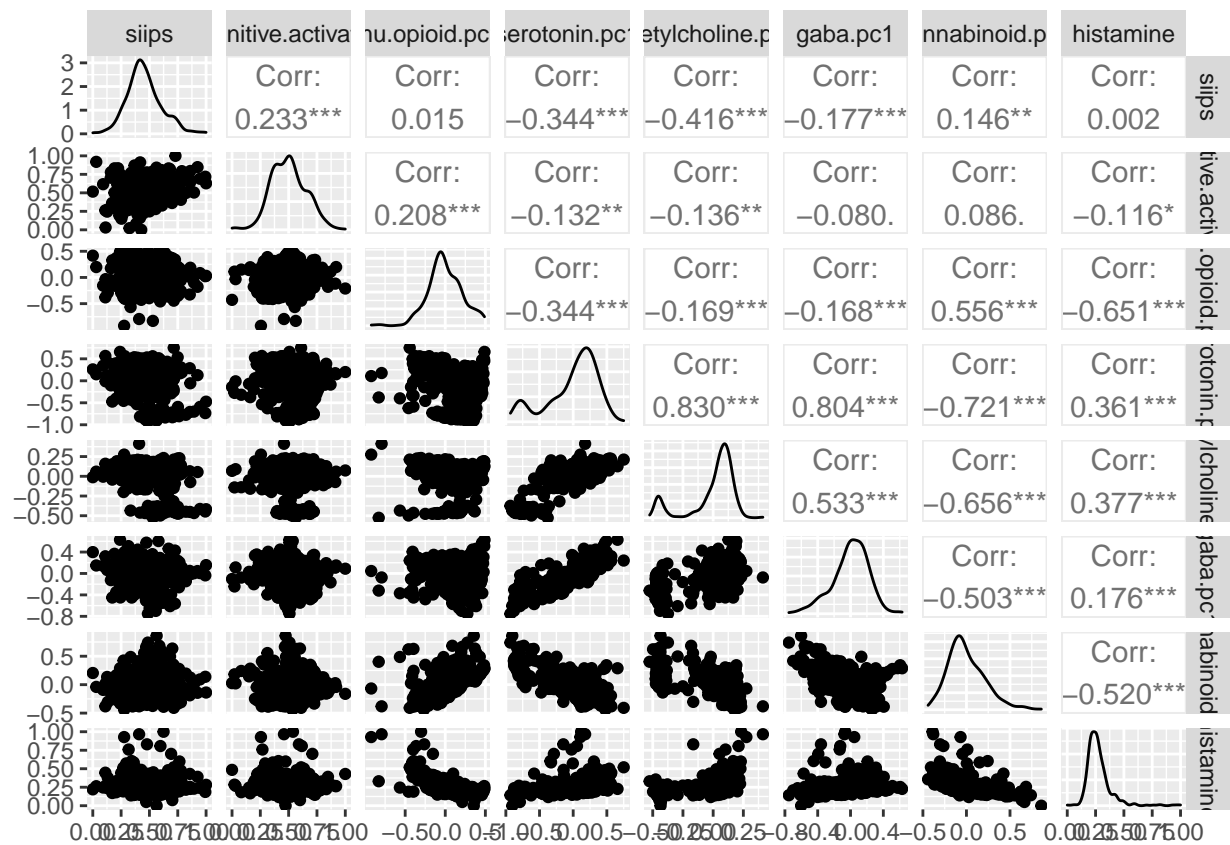
Feature selection

using in-sample criteria to maximize R^2

```
xs = as.matrix(neuromaps_pca[, -1])
all_subsets = leaps(xs, neuromaps_pca$siips, method = "adjr2")
col_idx = all_subsets$which[which.max(all_subsets$adjr2), ]
neuromaps_subset = data.frame(siips = neuromaps_pca$siips, xs[, col_idx])
```

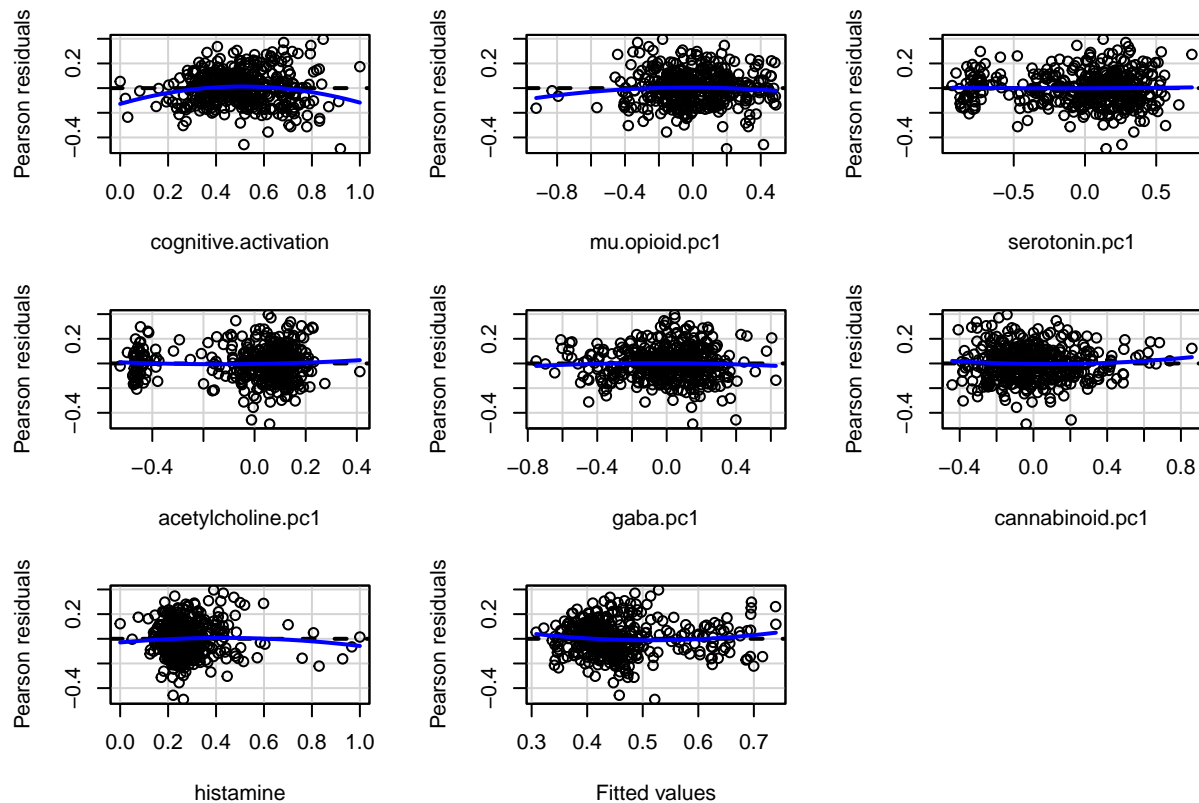
Pairs Plot

```
ggpairs(neuromaps_subset)
```



Naive model

```
ols1 = lm(siips ~ ., data = neuromaps_subset)
residualPlots(ols1) # nonlinearity
```



```
##                               Test stat Pr(>|Test stat|)
## cognitive.activation         -3.0341      0.002557 **
## mu.opioid.pc1                -1.6227      0.105369
## serotonin.pc1                 0.2382      0.811865
## acetylcholine.pc1            0.7013      0.483506
## gaba.pc1                     -0.6238      0.533089
## cannabinoid.pc1              1.0840      0.278982
## histamine                    -1.1978      0.231653
## Tukey test                    1.9282      0.053836 .
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
vif(ols1) # collinearity
```

```
## cognitive.activation      mu.opioid.pc1      serotonin.pc1
##           1.088422          2.838364          10.478633
##   acetylcholine.pc1      gaba.pc1      cannabinoid.pc1
##           5.358733          3.783645           3.015873
##           histamine
##           2.103284
```

```
ncvTest(ols1) # heteroscedasticity
```

```
## Non-constant Variance Score Test
## Variance formula: ~ fitted.values
## Chisquare = 8.132561, Df = 1, p = 0.0043477
```

Addressing nonlinearity: quadratic terms

Addressing collinearity: removing serotonin

```
ols2 = update(ols1, ~ . - serotonin.pc1 + I(cognitive.activation^2) + I(mu.opioid.pc1^2))
```

```
anova(ols1, ols2) # confirm ols2 is a better model
```

```
## Analysis of Variance Table
```

```
##
```

```
## Model 1: siips ~ cognitive.activation + mu.opioid.pc1 + serotonin.pc1 +  
##           acetylcholine.pc1 + gaba.pc1 + cannabinoid.pc1 + histamine
```

```
## Model 2: siips ~ cognitive.activation + mu.opioid.pc1 + acetylcholine.pc1 +  
##           gaba.pc1 + cannabinoid.pc1 + histamine + I(cognitive.activation^2) +  
##           I(mu.opioid.pc1^2)
```

```
##   Res.Df    RSS Df Sum of Sq      F   Pr(>F)
```

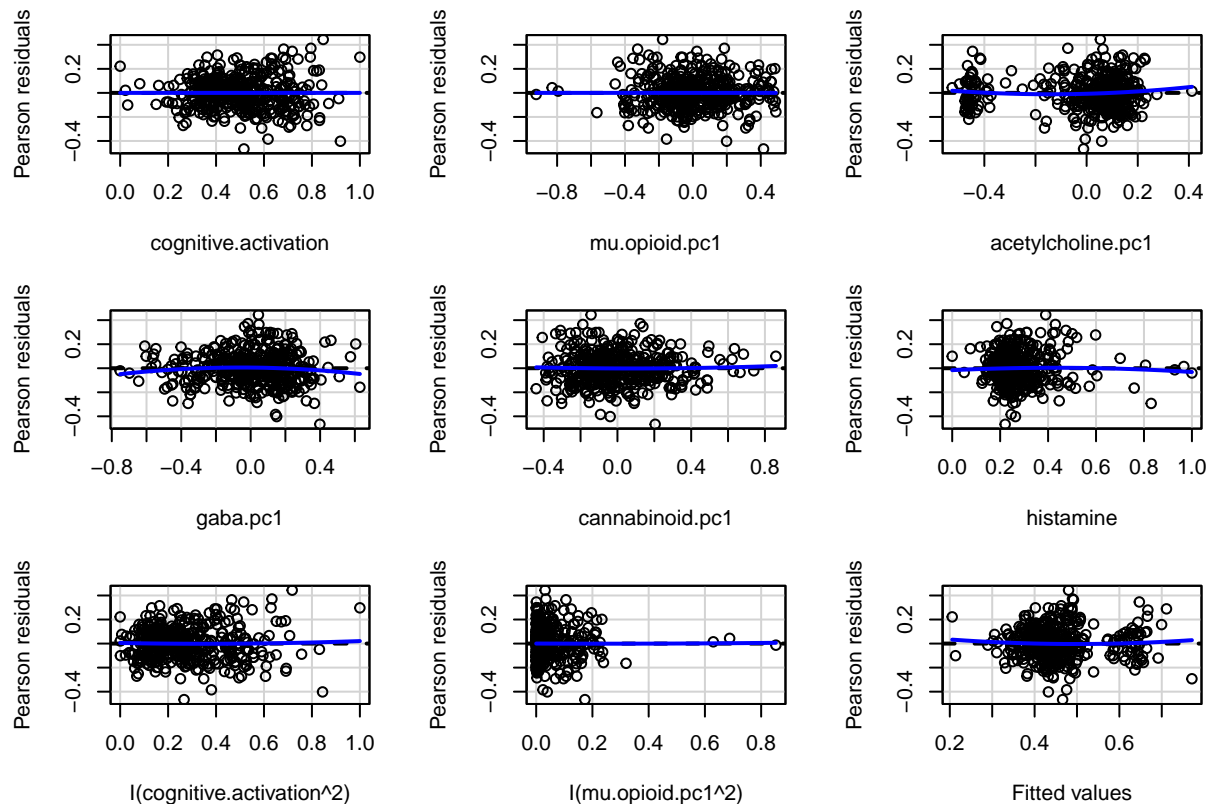
```
## 1      436 7.6417
```

```
## 2      435 7.4994  1   0.14233 8.2556 0.004261 **
```

```
## ---
```

```
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
residualPlots(ols2) # fixes nonlinearities
```



```
##                                     Test stat Pr(>|Test stat|)
```

## cognitive.activation	-1.6190	0.1062
## mu.opioid.pc1	0.5513	0.5817
## acetylcholine.pc1	1.3049	0.1926
## gaba.pc1	-1.5065	0.1327
## cannabinoid.pc1	0.4015	0.6883
## histamine	-0.6729	0.5013
## I(cognitive.activation^2)	0.6000	0.5488

```
## I(mu.opioid.pc1^2)          0.0850          0.9323
## Tukey test                  0.8646          0.3873
```

```
ncvTest(ols2) # heteroscedasticity still an issue
```

```
## Non-constant Variance Score Test
## Variance formula: ~ fitted.values
## Chisquare = 8.027564, Df = 1, p = 0.0046071
```

Check to see if errors are autocorrelated

```
resid = residuals(ols2)
cor(resid[-1], resid[-length(resid)])
```

```
## [1] 0.04566048
```

```
durbinWatsonTest(ols2) # GLS model doesn't seem nessessary
```

```
## lag Autocorrelation D-W Statistic p-value
## 1 0.04565254 1.908337 0.354
## Alternative hypothesis: rho != 0
```

Bootstrap to re-estimate standard errors and t-values, under heteroscedasity

```
n_bootstraps = 1000
n_samples = nrow(neuromaps_subset)
ols_coefs = ols2$coefficients
boot_coefs = matrix(0, n_bootstraps, length(ols_coefs))

for (i in 1:n_bootstraps) {
  idx = sample(1:n_samples, replace = TRUE)
  m = update(ols2, ~., data = neuromaps_subset[idx, ])
  boot_coefs[i, ] <- m$coefficients
}

stderr = apply(boot_coefs, 2, sd)
ci = apply(boot_coefs, 2, quantile, probs = c(0.025, 0.975))
tvals = ols_coefs / stderr
pvals = 2 * pt(-abs(tvals), df = nrow(neuromaps_subset) - length(ols_coefs))
```

Compiling bootstrap results

```
boot_summary = data.frame(
  "Estimate" = ols_coefs,
  "Std Error" = stderr,
  "t value" = tvals,
  "p value" = pvals
)
```

Compare summary tables

```
summary(ols2)$coefficients |> round(3)
```

##	Estimate	Std. Error	t value	Pr(> t)
## (Intercept)	0.119	0.056	2.136	0.033
## cognitive.activation	0.795	0.196	4.057	0.000
## mu.opioid.pc1	0.093	0.046	2.031	0.043
## acetylcholine.pc1	-0.469	0.051	-9.110	0.000
## gaba.pc1	0.045	0.035	1.276	0.203
## cannabinoid.pc1	-0.090	0.058	-1.540	0.124
## histamine	0.420	0.089	4.709	0.000
## I(cognitive.activation^2)	-0.616	0.186	-3.311	0.001
## I(mu.opioid.pc1^2)	-0.205	0.099	-2.084	0.038

```
boot_summary |> round(3)
```

##	Estimate	Std.Error	t.value	p.value
## (Intercept)	0.119	0.065	1.829	0.068
## cognitive.activation	0.795	0.253	3.138	0.002
## mu.opioid.pc1	0.093	0.046	2.007	0.045
## acetylcholine.pc1	-0.469	0.055	-8.502	0.000
## gaba.pc1	0.045	0.037	1.221	0.223
## cannabinoid.pc1	-0.090	0.052	-1.731	0.084
## histamine	0.420	0.093	4.511	0.000
## I(cognitive.activation^2)	-0.616	0.261	-2.358	0.019
## I(mu.opioid.pc1^2)	-0.205	0.083	-2.488	0.013

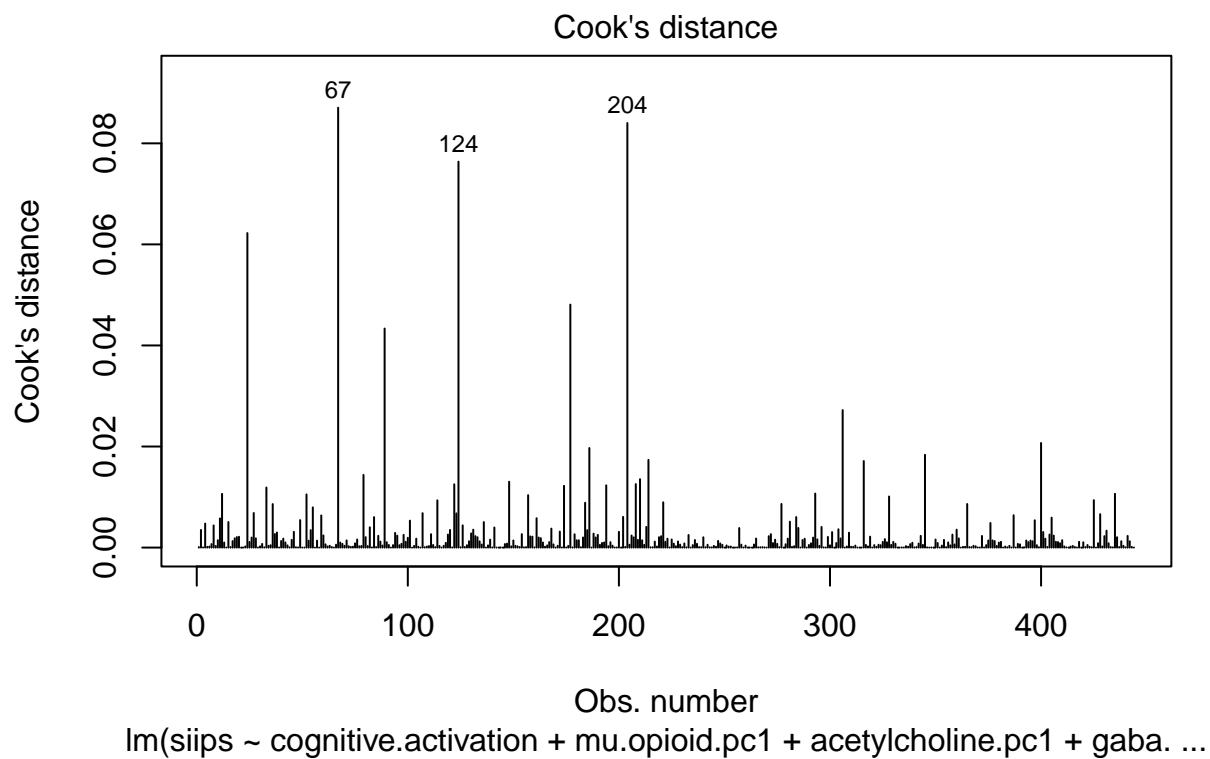
Compare confidence intervals

```
confint(ols2) |>  
  cbind(t(ci)) |>  
  round(3)
```

##	2.5 %	97.5 %	2.5%	97.5%
## (Intercept)	0.010	0.228	-0.025	0.232
## cognitive.activation	0.410	1.180	0.361	1.371
## mu.opioid.pc1	0.003	0.183	0.009	0.189
## acetylcholine.pc1	-0.570	-0.367	-0.572	-0.363
## gaba.pc1	-0.024	0.115	-0.028	0.116
## cannabinoid.pc1	-0.204	0.025	-0.196	0.004
## histamine	0.244	0.595	0.266	0.626
## I(cognitive.activation^2)	-0.982	-0.250	-1.195	-0.163
## I(mu.opioid.pc1^2)	-0.399	-0.012	-0.385	-0.047

Outliers

```
plot(ols2, 4)
```

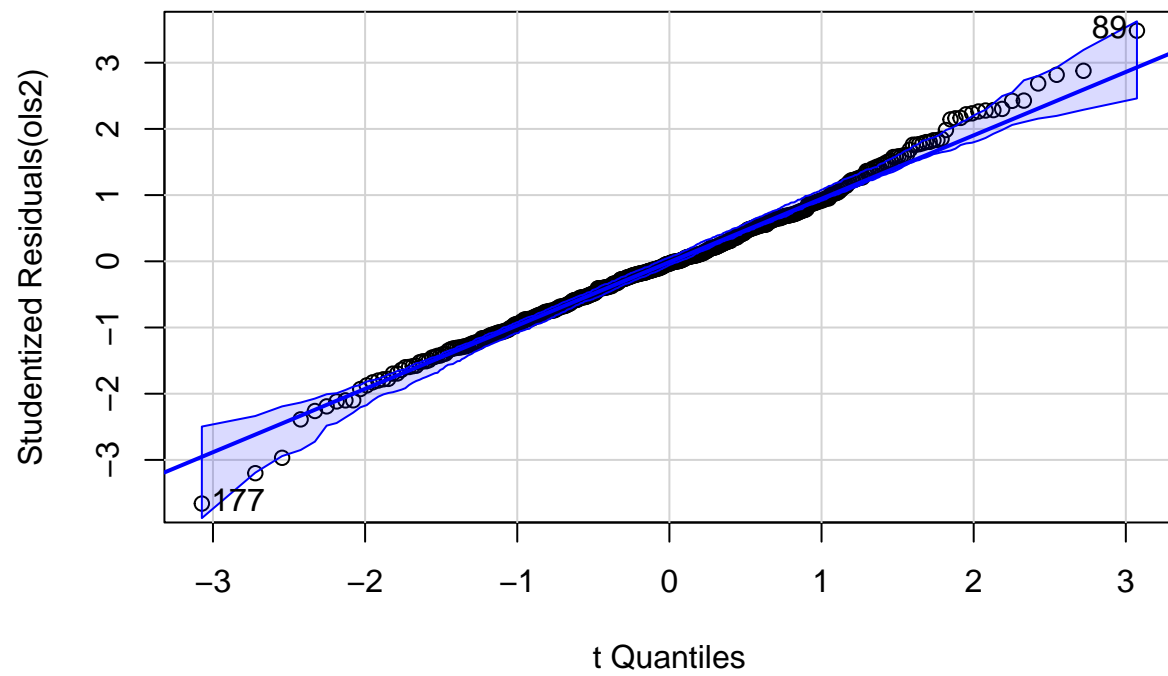


```
outlierTest(ols2)
```

```
## No Studentized residuals with Bonferroni p < 0.05
## Largest |rstudent|:
##      rstudent unadjusted p-value Bonferroni p
## 177 -3.659496      0.00028381      0.12601
```

Normality of errors

```
qqPlot(ols2)
```

[1] 89 177