

# **ScienceDirect**



### **Review Article**

# Calcium-mediated mitochondrial energy deficiency in Parkinson's and Alzheimer's diseases: Insights from computational modelling

Valérie Voorsluijs<sup>1</sup> and Alexander Skupin<sup>2,3,4,a</sup>

Alzheimer's and Parkinson's diseases are the most prevalent neurodegenerative disorders worldwide and are characterised by progressive cognitive and functional impairments caused by neuronal loss. Energy deficiency is a predominant hallmark of their pathophysiology and plays a central role in the development of the disease, notably by mitochondrial dysfunction enhancing protein aggregation and oxidative stress which trigger subsequently immune responses and neuronal loss. Quantifying this energetic deficiency and identifying specific causative mechanisms from the complex network of interacting metabolic and regulatory pathways at play is rather challenging, where integrative mathematical modelling represents a powerful tool to support these investigations. Here, we review the latest developments in integrative modelling in brain bioenergetics in relation to Alzheimer's and Parkinson's diseases where we focus on the regulatory role of Ca<sup>2+</sup> signalling. Finally, we discuss recent challenges and future directions to improve the current understanding of the energy-deficiency theory of neurodegeneration.

#### Addresses

Luxembourg

- <sup>1</sup> Cancer Metabolism Group, Department of Cancer Research, Luxembourg Institute of Health, 6a rue Nicolas-Ernest Barblé, Luxembourg, L-1210, Luxembourg
- <sup>2</sup> Luxembourg Centre for Systems Biomedicine, University of Luxembourg, 6 avenue du Swing, Belvaux, L-4367, Luxembourg
   <sup>3</sup> Department of Physics and Materials Science, University of Luxembourg, 162 A avenue de la Faïencerie, Luxembourg, L-1511,
- <sup>4</sup> Department of Neurosciences, University of California San Diego, 9500 Gilman Drive, San Diego, 92093, CA, USA

Corresponding author: Skupin, Alexander (alexander.skupin@uni.lu) a http://wwwen.uni.lu/lcsb/research/integrative\_cell\_signalling.

#### Current Opinion in Systems Biology 2025, 40:100539

This review comes from a themed issue on **Mathematical Modelling** (2023)

## Edited by Jana Wolf and Kevin Thurley

For complete overview of the section, please refer the article collection - Mathematical Modelling (2023)

Available online 10 January 2025

# https://doi.org/10.1016/j.coisb.2024.100539

2452-3100/© 2025 The Authors. Published by Elsevier Ltd. This is an open access article under the CC BY license (http://creativecommons.org/licenses/by/4.0/).

#### Keywords

Computational modelling, Neurodegeneration, Calcium, Energy deficiency, Mitochondria, Metabolic reprogramming, Translational research.

#### Introduction

Alzheimer's disease (AD) and Parkinson's disease (PD) are recognised as the most prevalent neurodegenerative disorders worldwide. Symptomatic relief can be achieved with the currently available treatments, but a sustainable cure for those diseases has so far not emerged [1,2]. Both diseases are characterised by cognitive decline and functional impairments caused by neuronal death in specific brain regions, and are associated with protein accumulation in the brain, neuroinflammation, brain atrophy, increased oxidative stress, mitochondrial dysfunction, remodelling of glucose metabolism and dysregulation of Ca<sup>2+</sup> dynamics [1,2].

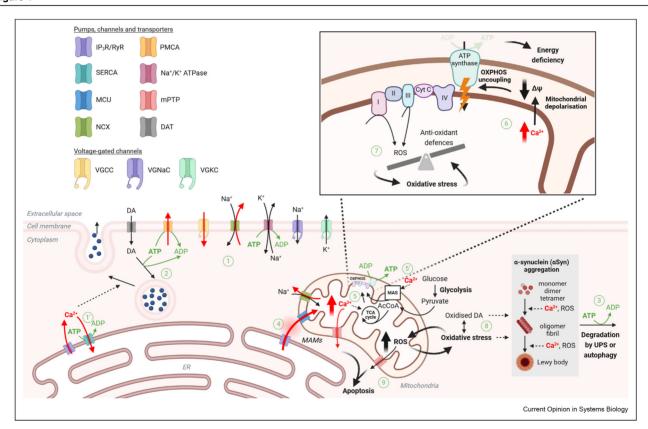
It is now widely accepted that the accumulation of  $\alpha$ synuclein in PD or of amyloid  $\beta$  (A $\beta$ ) and neurofibrillary tangles composed of hyperphosphorylated tau proteins in AD leads to various anomalies and eventually to neurodegeneration [2,3]. The characteristic motor symptoms of PD mainly result from the death of dopaminergic neurons in the substantia nigra pars compacta (SNc) [2] whereas memory loss in AD is attributed to impairments in synaptic transmission and notably to the degeneration of cholinergic neurons in the basal forebrain [1]. Accumulating evidence also points to a remodelling of cellular interactions across the brain cell network in both AD and PD, and highlights the potential contribution of non-neuronal cells such as astrocytes and microglia [4–6]. These glial cells secrete various molecules (gliotransmitters, cytokines, chemokines etc.) that not only regulate synaptic transmission and plasticity, but also control brain inflammatory responses [5,7]. Anomalous release/uptake of these molecules, dysfunctional signal transduction or sustained release of pro-inflammatory cytokines, with their potential amplification by excess protein aggregates, constitute major causes of neurotoxicity and neuronal death [5,8].

Nevertheless, the exact sequence of events driving neurodegeneration remains elusive. Aging, genetic mutations and exposure to environmental toxins cause many disturbances in physiology that could act upstream of protein deposition [9]. Among those, impaired bioenergetics, characterised by adenosine triphosphate (ATP) deficiency, stands out as another early and robust hallmark of neurodegeneration [9]. Various signalling and housekeeping processes that are essential to proper synaptic function and neuron homeostasis require ATP [10,11]. For instance, the maintenance of the resting membrane potential and the encapsulation of neurotransmitters in vesicles are mediated by ATPases. Alterations in different

metabolic, signalling or transport pathways, notably reducing glucose uptake and mitochondrial efficiency, compromise ATP production while maintaining (or even increasing) the neuronal energy demand (Figure 1). Moreover, neuroinflammation triggers a metabolic rewiring in microglia [12,13] that limits glucose and lactate availability in the tissue and could potentially induces ATP deficiency in neurons [14]. Targetting the aforementioned pathways (Figure 1) to prevent ATP deficiency thus appears as a promising strategy for therapeutic intervention [15].

However, understanding the mechanisms of disease onset and progression appears as an intractable problem if

Figure 1



Energetic and Ca2+ contributions to the onset and progression of PD. The homeostasis of SNc dopaminergic neurons is associated with a high energetic cost due to the following main mechanisms. (1)/(1') Action potentials are accompanied by large Ca2+ entry through VGCC and recovery is mediated by ATPases (PMCA, SERCA, Na\*/K\* ATPase). (2) DA encapsulation in vesicles is enabled by the vesicular monoamine transporter, a V-ATPase. The vesicle recycling (not shown) also involves a V-ATPase. (3) Damaged protein disposal mechanisms mediated by UPS and lysosomes require ATP (ubiquitination, ATP-dependent proteasome and lysosomal V-ATPase). (4) The Ca<sup>2+</sup> influx induced by VGCC is amplified by IP<sub>3</sub>Rs and RyRs through a process of Ca<sup>2+</sup>-induced Ca<sup>2+</sup> release and readily taken up by mitochondria due to the close proximity between these channels and the MCU in MAMs. (5)/(5') High Ca<sup>2+</sup> level stimulates mitochondrial NADH availability by activating the TCA cycle (5) or via the MAS (5'). NADH enters the ETC which establishes the proton motive force driving the synthesis of ATP by ATP synthase. The intense stimulation of OXPHOS tends to build up a basal oxidative stress due to the production of ROS. (6) Under some circumstances (e.g. PD-related mutations, mitochondrial dysfunction etc.), altered mitochondrial Ca<sup>2+</sup> exchanges lead to Ca<sup>2+</sup> overload, which decreases the mitochondrial membrane potential and uncouples the ETC from ATP production. The disruption of mitochondrial ATP synthesis is a major cause of energy deficiency. (7) ROS production increases due to the overstimulation of the ETC by Ca2+ and the anti-oxidant defence system becomes overwhelmed. The resulting oxidative stress is self-amplified by the mitochondrial damage caused by ROS, which further alters the activity of the ETC. (8) The leakage of ROS to the cytosol activates dopamine oxidation. The presence of oxidised dopamine as well as high ROS and Ca<sup>2+</sup> levels favours the aggregation of αSyn into oligomers and Lewy bodies. Importantly, energy deficiency prevents the alleviation of these stress conditions by dopamine encapsulation (2), Ca2+ removal (1)/(1') or protein degradation (3), which are all ATP-dependent. (9) Ca<sup>2+</sup> and ROS accumulation in mitochondria finally induces the opening of mPTP, which triggers an apoptotic signalling cascade (caspase) leading to neuronal death. (Red and black dashed arrows represent Ca2+ fluxes and direct activation of another process, respectively.)

we consider the highly intricate network of intra- and intercellular regulation pathways controlling the ATP level (Figure 1) [15]. Mathematical modelling has emerged as a powerful tool to address those challenges by providing modular and integrative approaches [16] where the response of a core brain energy model [17] to individual disease-related mechanisms [18] can be analysed. Here, we review the developments from the past 10 years in integrative modelling of brain cellular bioenergetics and focus on the mechanistic models capturing AD- and PD-related alterations with an emphasis on Ca<sup>2+</sup> homeostasis. Models of astrocyte and neuron metabolism [19], of AD and PD pathogenesis [20,21] or of mitochondria in health and neurodegeneration [22] have been extensively reviewed elsewhere.

After presenting the major classes of models with their scope and limitations, we elaborate on how to integrate them in frameworks to more systematically quantify energetic damage in neurodegeneration or envision multiscale and translational models.

# Overview of recent mechanistic models of brain cell energetics PD models

The shape of dopaminergic neurons in the SNc has been proposed to lead to their intrinsic vulnerability, due to the mismatch between their energy budget, on the one hand, and the energetically demanding large number of synaptic connections and size of their unmyelinated axonal architecture, on the other hand [23,24]. Morphological and electrophysiological models of those cells have shown that the propagation and recovery from an action potential represent extremely large energy demands that could probably not be balanced in unhealthy cells with dysfunctional metabolism [25,26].

The autonomous pacemaking of SNc dopaminergic neurons, i.e. their ability to sustain continuous spiking without synaptic excitatory stimulus, leads to the repetitive opening of voltage-gated Cav1 Ca<sup>2+</sup> channels (VGCC L-type Ca<sup>2+</sup> channels) and to high-amplitude oscillations in Ca<sup>2+</sup> concentration [27,23,24]. The low cytosolic Ca<sup>2+</sup> buffering capacity of these neurons is a double-edge sword. Low buffering favours Ca<sup>2+</sup> accumulations that promote the aggregation of α-synuclein [27,23,24] or trigger apoptotic signals via the opening of the mitochondrial permeability transition pore (mPTP) [23]. However, high-amplitude Ca<sup>2+</sup> spikes have a major impact on the cellular energy balance, not only due to the energetic cost of Ca<sup>2+</sup> extrusion, but also through the activatory effect of Ca<sup>2+</sup> on OXPHOS [28,29] (Figure 1). This regulation proceeds either through direct activation of mitochondrial dehydrogenases of the tricarboxylic acid (TCA) cycle, which relies on Ca<sup>2+</sup> transport via the mitochondrial Ca<sup>2+</sup> uniporter (MCU) and enhances the flux of the electron transport (ETC) chain, or is mediated by the Ca<sup>2+</sup>-regulated Aralar/Malate-Aspartate

shuttle (MAS), involved in NADH transport from the cytosol to mitochondria [30,31].

Some PD-related mutations affect the expression of mitochondrial  ${\rm Ca}^{2+}$  transporters, leading to perturbed  ${\rm Ca}^{2+}$  exchanges,  ${\rm Ca}^{2+}$  overload, mitochondrial depolarisation and inefficient coupling between electron transfer and ATP synthesis. Soman et al. have computationally predicted that decreasing the flux through MCU prevents mitochondrial Ca<sup>2+</sup> overload and restores mitochondrial respiration in neurons carrying a PTEN-induced putative kinase 1 (PINK1) mutation, which was also confirmed experimentally [32]. Silencing or pharmacological inhibition of MCU rescued dopaminergic neurons of zebrafish embryos affected by PINK1 loss-of-function [32]. Their non-spatial computational model captured the cross-talk between Ca<sup>2+</sup> dynamics and metabolism by key steps of cytosolic and mitochondrial glucose metabolism, regulated by Ca<sup>2+</sup> exchanges between the cytosol, the endoplasmic reticulum and mitochondria.

Notably, Soman et al. neglected the neuronal membrane electrophysiology and the level of reactive oxygen species (ROS) was not measured nor modelled [32]. However, the excessive production of ROS is a direct consequence of disrupted mitochondrial respiration and the cause of serious cellular damage, ranging from oxidation of essential biomolecules (proteins, lipids, DNA) to impairments in mitochondrial function and quality control [23]. In fact, the chronic exposure of SNc dopaminergic neurons to high ROS levels also contributes to their vulnerability [23]. Indeed, the feedforward activation of OXPHOS by Ca<sup>2+</sup>, although key in the maintenance of autonomous pacemaking [30], tends to produce high levels of ROS in physiological conditions and thereby generates a basal oxidative stress [33]. Depending on the intracellular redox conditions, the tight interplay between Ca<sup>2+</sup> and ROS promotes beneficial cell signalling or amplifies abnormal behaviours in pathological conditions [34-36]. Whether the loss of these optimal intracellular redox conditions triggers PD onset remains overall elusive [35].

Models capturing the Ca<sup>2+</sup>-dependent electrophysiology, redox balance and metabolism of dopaminergic SNc neurons are thus essential to estimate the actual gap between energy supply and demand in those cells. A recent model supporting an energy-deficiency hypothesis for PD pathogenesis [37] constitutes a promising baseline. This model accounts for the dynamics of membrane potential and ionic membrane currents, Ca<sup>2+</sup> dynamics, glucose metabolism, dopamine metabolism and (Ca<sup>2+</sup>-dependent) release, ER-stress-induced apoptotic pathways as well as pathways specific to the pathophysiology of PD, adapted from the pioneering work of Cloutier and Wellstead [18]. Briefly, this PD module captures α-synuclein metabolism, protein degradation pathways, oxidative stress and ROS clearance through the anti-oxidative pathway, with a feedback loop between ROS and α-synuclein dynamics. Energetic costs, expressed in terms of ATP consumption rate, are associated to the propagation and recovery from action potentials, synaptic vesicle management, Ca<sup>2+</sup> pumping and degradation of damaged proteins.

Hypoglycemic and hypoxic conditions induce energy deficiency, i.e. low ATP availability, which limits the degradation of misfolded and aggregated α-synuclein, favours the accumulation of ROS and Ca<sup>2+</sup> and induces a vicious cycle between ROS, Ca<sup>2+</sup>, dopamine and αsynuclein dynamics (Figure 1). Experimental evidence of this feedback cycle (comprehensively reviewed elsewhere [38]) supports a multiple hit hypothesis for PD. Although data do not point directly to hypoxia or hypoglycemia as the trigger of the pathological cascade, these conditions might mimic critical upstream perturbations, such as mitochondrial oxidative stress [39]. Of note, similar energy deficiency issues were reported with a simpler computational model of glucose and redox metabolism, and validated against metabolomics measurements from a Parkin knockout mice brain [40]. While Muddapu et al. focussed on energy deficiency, their mathematical model contains the necessary components to formulate different starting hypotheses (e.g. on redox conditions, status of antioxidant defences, mitochondrial Ca<sup>2+</sup> load, status of mitochondrial respiration etc.), which could help clarify the chronology and interdependencies of dysfunctions in the pathogenesis of PD.

This integrative model could be extended with the Ca<sup>2+</sup> regulation of mitochondrial metabolism to account for Ca<sup>2+</sup>-induced metabolic adaptations in response to an action potential. Further model refinements could include the MAS, the Ca<sup>2+</sup> regulation of mPTP and the modelling of mitochondria-associated membranes (MAMs) [41]. The spatial extent of those cytosolic domains, where the ER is in close interaction with the outer mitochondrial membrane, is modified in PD and in AD, thereby altering Ca<sup>2+</sup> exchanges and, possibly, mitochondrial ATP synthesis [42]. Finally, introducing the interactions with other cells such as astrocytes and microglia to reproduce a tri- or quad-partite synapse would improve the physiological relevance of the model, as discussed in Section From sub-organellar to systems scales.

#### **AD models**

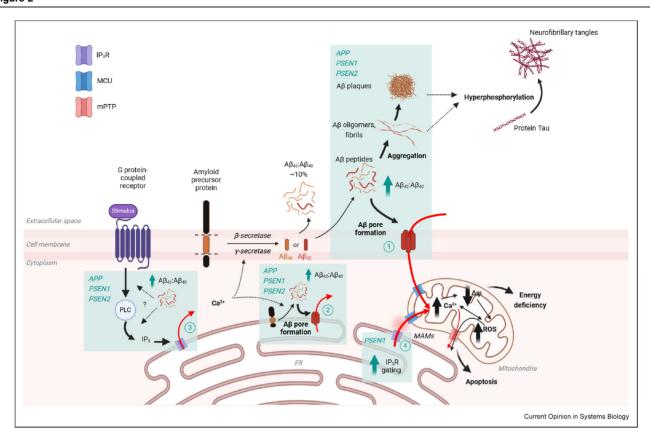
Familial and sporadic AD are both characterised by abnormal A $\beta$  accumulation [3]. The cleavage of amyloid precursor protein (APP) by the  $\beta$ - and  $\gamma$ -secretase complexes releases A $\beta$  peptides and takes place predominantly in the plasma membrane, ER and Golgi network [43]. In familial AD, mutations in genes coding

for the APP or part of the  $\gamma$ -secretase complex enhance the formation of protein deposits, including A $\beta$  plaques and neurofibrillary tangles [1,3] (Figure 2).

These mutations are responsible for many other pathological phenomena including impaired Ca<sup>2+</sup> signalling [3,44]. Recent therapeutic strategies consider the socalled "Ca<sup>2+</sup> hypothesis" and its tight connection to energy deficiency as a complementary view to the canonical amyloidogenic pathway [45]. Perturbations in Ca<sup>2+</sup> dynamics arise very early in AD development, preceding the appearance of extracellular amyloid deposits and resulting most likely from intracellular AB dyshomeostasis [46,43], although amyloid-independent mechanisms have also been identified [47] (Figure 2). Of note, a PSEN1 mutation seems to increase the opening probability of IP<sub>3</sub>Rs in an Aβ-independent way [48]. Intracellular Aβ increases cytosolic Ca<sup>2+</sup> levels by inducing ER release through  $A\beta_{42}$  pores formed in the ER membrane [49] or stimulation of IP<sub>3</sub>Rs by Aβenhanced production of IP<sub>3</sub> [50,49], which perturbs the Ca<sup>2+</sup> fluxes of other intracellular organelles such as mitochondria [51]. Additionally, pore-forming assemblies of extracellular A\beta in the plasma membrane (PM) enables large influx of extracellular Ca<sup>2+</sup> [52]. Since Aβ production is itself modulated by Ca<sup>2+</sup>, AB and Ca<sup>2+</sup> are mutually activating their production and the resulting feedforward loop eventually causes neurodegeneration [44]. Moreover, mitochondrial AB accumulation decreases the activity of complexes III and IV in the ETC, leading to mitochondrial dysfunction and ROS accumulation [43].

In computational models, these AD-related perturbations can be incorporated in the description of cytosolic Ca<sup>2+</sup> dynamics (Table 1). Several gating models, predicting the opening probability of IP<sub>3</sub>Rs and the associated Ca<sup>2+</sup> release from the ER, have been established based on experimental Ca<sup>2+</sup> traces. Regardless of the exact mechanism rising cytosolic Ca<sup>2+</sup>, cytosolic Ca<sup>2+</sup> traces feed a bioenergetic model that predicts the evolution of key mitochondrial variables such as Ca<sup>2+</sup>, ATP, NADH and ROS concentrations, and mitochondrial membrane potential. Cortassa's bioenergetic model [53,54] is widely used and accounts for the production of ATP through the TCA cycle and OXPHOS, and includes details of enzyme regulation - notably, the regulation of isocitrate dehydrogenase and α-ketoglutarate dehydrogenase by Ca<sup>2+</sup>. This model also incorporates the anaplerotic reaction catalysed by aspartate transaminase, ROS generation by the respiratory chain and ROS scavenging. Overall, the computational models indicate that abnormal Ca<sup>2+</sup> elevations affect the energy metabolism and ROS production by inducing mitochondrial Ca<sup>2+</sup> overload and mitochondrial depolarisation, which in turn decreases the proton motive force driving the synthesis of ATP [55,51,56,57]

Figure 2



Ca2+ fluxes directly or indirectly associated with AD-related mutations leading to energy deficiency and apoptosis. The canonical pathway of APP cleavage by  $\beta$ - and  $\gamma$ -secretases yields a mixture of A $\beta$  monomers of 40 or 42 residues with an A $\beta_{42}$ :A $\beta_{40}$  ratio of about 10 %. In the presence of autosomal dominant inherited mutations of APP PSEN1 or PSEN2 genes, APP cleavage generates a higher fraction of ABa2 which tend to aggregate into oligomers, fibrils and plaques. The metabolism of protein tau is altered by these aggregates, which leads to the assembly of neurofibrillary tangles. These mutations also remodel Ca<sup>2+</sup> by favouring the formation of pores in the plasma membrane (1) or ER membrane (2), by enhancing IP<sub>3</sub>-mediated stimulation of IP<sub>3</sub>Rs (3) or increasing the open probability of IP<sub>3</sub>Rs (4). Mitochondria take up these excess amounts of Ca<sup>2+</sup>, which decreases the mitochondrial membrane potential ( $\Delta\psi$ ). ROS production is enhanced by Ca<sup>2+</sup> overload and decoupling of OXPHOS, which decreases ATP synthesis and leads to energy deficiency - the details of the energetic perturbation cascade can be found in Figure 1. Finally, high intramitochondrial levels of ROS and Ca<sup>2+</sup> trigger the opening of the mitochondrial transition pore (mPTP) and the release of apoptotic signals. (Red and black dashed arrows represent Ca<sup>2+</sup> fluxes and direct activation of another process, respectively.)

(Figure 2). Additionally, the model of Toglia and Ullah shows that the spatial configuration of the MAMs modulates the opening probability of the mPTP, which mitochondrial also controls metabolism apoptosis [51].

Ca<sup>2+</sup> and metabolic remodelling affects non-neuron cells, as exemplified by the reactive astrocytes and activated microglia observed in the neighbourhood of plaques and tangles [7]. Under some conditions, astrocytes provide energy support to neurons in the form of lactate supply (astrocyte-neuron lactate shuttle) [15]. As shown with a spatio-temporal model of glucose metabolism, reactive astrocytes exposed to metabolic AD-related perturbations exhibit globally reduced ATP:ADP ratio and lactate export rate, which points to ATP deficiency and poor energetic support to neurons [61]. Interestingly, the morphological adaptation of the reactive astrocyte could locally mitigate the effects of AD-associated metabolic alterations in these cells due to limited transport in the ramified processes [61]. This hypothesis should be further investigated with a refined energy metabolism model and in a more specific context (synaptic transmission, neuroinflammation,  $Ca^{2+}$ signalling etc.) since the reactive transformation of astrocytes is shaped by the nature of the trigger [62]. The comprehensive and astrocyte-specific model of Latulippe et al. could for instance be incorporated to account for the  $Ca^{2+}$ -A $\beta$  interplay [59]. How metabolites are spatially distributed through the cell or within brain tissue and how intercellular interactions perturb these profiles remain largely open questions that could be addressed with metabolic models of high spatiotemporal resolution.

Def	A.D. manuturulu attau	Cytosolic Ca <sup>2+</sup>	Matelani	olism.	On all all	Other market to the state of th
Ref	AD perturbation	handling	Metabolism	Energy measure	Spatial model	Other model assumptions
Mak et al.2015 [58]	Enhanced IP <sub>3</sub> R gating by PS mutations	IP <sub>3</sub> R gating model fitted to exp. data from Sf9 cells expressing mutant PS1	-	-	3D reaction-diffusion model	Opening of IP3Rs (stochastic) and Ca <sup>2+</sup> release (deterministic). Ca <sup>2</sup> buffering by dye (deterministic).
Toglia et al. 2016 [55]	Enhanced IP <sub>3</sub> R gating by PS mutations	Exp. Ca <sup>2+</sup> traces from lymphoblasts of AD patients expressing mutant PS1	TCA cycle, OXPHOS, ROS [53,54]	[ATP]	-	<del>-</del>
Toglia et al. 2016 [51]	Enhanced IP <sub>3</sub> R gating by PS mutations	IP <sub>3</sub> R gating model and parameters from Mak et al. 2015 [58]	TCA cycle, OXPHOS, ROS [53,54]	[ATP]	Ca <sup>2+</sup> in MAMs: reaction-diffusion model (Ca <sup>2+</sup> flux through IP <sub>3</sub> Rs + endogeneous buffering). Whole-cell model: non-spatial. Mitochondrial Ca <sup>2+</sup> fluxes (MCU, NCX, mPTP) depend on Ca <sup>2+</sup> level in MAMs	_
Toglia et al. 2018 [56]	${\rm A}eta_{ m 42}$ -enhanced ${\rm IP_3}$ production	IP <sub>3</sub> R gating and whole-cell models fitted to exp. data from <i>Xenopus</i> oocytes	-	[ATP]	Fitting of Ca <sup>2+</sup> traces: reaction-diffusion model. Mitochondrial metabolism: non-spatial.	Ca <sup>2+</sup> exchanges with ECS neglected. Exogenous cytosolic Ca <sup>2+</sup> buffering.
Latulippe et al. 2018 [59]	$A\beta$ -enhanced $Ca^{2+}$ entry. ( $A\beta$ pores in PM) and sensitivity of RyR	IP <sub>3</sub> Ř gating and whole-cell model	-	-	_	Astrocyte-specific. Detailed model of cell membrane potential dynamics. Ca <sup>2+</sup> exchanges with ECS (PMCA, leak, VGCC). RyR Module for IP <sub>3</sub> metabolism Mitochondrial exchanges and metabolism neglected.
Minicucci et al. 2021 [60]	$A\beta$ -enhanced $IP_3$ production	Whole-cell model fitted to exp. data from <i>Xenopus</i> oocytes	TCA cycle, OXPHOS, ROS [53,54]	-	-	Module for IP <sub>3</sub> metabolism Mitochondrial exchanges and metabolism neglected. Ca <sup>2+</sup> exchanges with ECS neglected.
Pensalfini et al. 2022 [57]	Brain extract- enhanced IP <sub>3</sub> production	IP <sub>3</sub> R gating and whole-cell models fitted to exp. data from <i>Xenopus</i> oocytes (approach similar to Toglia et al. 2018)	-	[ATP]	Fitting of Ca <sup>2+</sup> traces: reaction-diffusion model. Mitochondrial metabolism: non-spatial.	Ca <sup>2+</sup> exchanges with ECS neglected.

6

Mathematical Modelling (2023)

Astrocyte-specific. Spatially localised. enzymes. Coarse-grained metabolic kinetics (e.g. no enzyme regulation, mass action law kinetics for entire complex pathways).	
Reaction-diffusion model. Includes exp. astrocytic morphologies.	
ATP:ADP, lactose export rate	
Glycolysis, TCA cycle, OXPHOS, LDH	
Morphological – changes in astrocytes (based on images from <i>post mortem</i> AD patient brains)	
Farina et al. 2023 [61]	

presenilin, RyR – ryanodine receptor, mPTP – mitochondrial transition pore, LDH – Lactate dehydrogenase, ECS – extracellular space, VGCC – voltage-gated Ca<sup>2+</sup> channel

# Challenges and future directions

### Towards a thermodynamically consistent description

In most models of brain energetics, the concept of energetic cost/production remains loosely defined. Absolute ATP concentration, ATP:ADP ratio, ATP consumption rate per surface or volume, or lactate export rate have been reported, which makes model comparison quite difficult. The energy systems approach proposed by Wellstead and Cloutier [16] paves the way to a more systematic approach. A next step would be to normalise those costs with respect to the basal energy input provided to cells in the form of oxygen, mitochondrial substrate, etc. The obtained thermodynamic efficiency would then become independent of the energy currency and capture the actual bioenergetic work performed by cells. Based on topological analysis and nonequilibrium thermodynamics, our recent study estimated the thermodynamic efficiency of mitochondrial metabolism regulated by Ca<sup>2+</sup> for different cell stimulation and mitochondrial substrate levels [63]. Using similar frameworks in bioenergetics would for example avoid bias in estimations of energy deficiency induced by the decreased food intake in patients with neurodegenerative diseases. More generally, a unifying bioenergetic approach is necessary to rationalise the mechanisms connecting metabolic rewiring to the energetic demands and expenses, which are ubiquitous in neurodegenerative processes.

# From sub-organellar to systems scales

In the reviewed kinetic models, most extracellular pathways as well as the dynamics of the intramitochondrial structures supporting ATP synthesis were overlooked. However, the internal structures of mitochondria, and more particularly cristae, can remodel in dependence on the matrix Ca<sup>2+</sup> content, which affects the ATP synthesis capacities [64]. More or less sophisticated models enable to investigate the mechanisms of mitochondrial network adaptation due to fusion, fission, fragmentation and migration [22], while accounting for the ATP-dependency of these processes [65] or providing spatially resolved predictions in line with experimental data [66]. Even though mitochondrial adaptation proceeds on a different timescale compared to carbon metabolism or Ca<sup>2+</sup> signalling (which is also a regulator of mitochondrial transport), a multiscale spatio-temporal framework could provide a more realistic description of energy delivery mechanisms. Intercellular exchanges, especially in the context of astrocytes and microglia activation, seem to significantly affect resource availability. With the growing interest for immunometabolism, the field of integrative bioenergetics should also witness the emergence of more models accounting for microglial involvement [14] and neuroinflammatory pathways.

Translational approaches based on mechanistic modelling remain scarce, but a recent attempt to connect molecular and cellular responses to motor skills succeeded in reproducing the symptomatic behaviours associated with PD and their improvement under medication with levodopa [67]. The model also offers clinical perspective by predicting the optimal level of levodopa to be administered in dependence on observed cell loss [67].

How impaired energetics and Ca<sup>2+</sup> signalling are linked and contribute to neurodegeneration becomes more and more evident, highlighting the therapeutic potential of interventions targetting Ca<sup>2+</sup> pumps and channels [45]. Nevertheless, many details still remain elusive and experiments guided by mechanistic modelling offer promising possibilities to unravel the underlying mechanisms of neurodegeneration.

# **Declaration of competing interest**

None.

## **Acknowledgements**

AS and VV acknowledge the support from the Luxembourg National Research Fund (FNR) through the ReFoRMCaS project (C23/BM/18093639/ReForMCaS). Figures created with BioRender.

### Data availability

No data was used for the research described in the article.

#### References

Papers of particular interest, published within the period of review, have been highlighted as:

- \* of special interest
- \*\* of outstanding interest
- Breijyeh Z, Karaman R: Comprehensive review on Alzheimer's disease: causes and treatment. Molecules 2020, 25:5789, https://doi.org/10.3390/molecules25245789.
- Bloem BR, Okun MS, Klein C: Parkinson's disease. Lancet 2021, 397:2284–2303, https://doi.org/10.1016/S0140-6736(21)00218-X.
- Hampel H, Hardy J, Blennow K, Chen C, Perry G, Kim SH, Villemagne VL, Aisen P, Vendruscolo M, Iwatsubo T, Masters CL, Cho M, Lannfelt L, Cummings JL, Vergallo A: The amyloid-β pathway in Alzheimer's disease. Mol Psychiatr 2021, 26: 5481-5503. https://doi.org/10.1038/s41380-021-01249-0.
- Liddelow SA: Modern approaches to investigating nonneuronal aspects of Alzheimer's disease. Faseb J 2019, 33: 1528–1535, https://doi.org/10.1096/fj.201802592.
- Miyazaki I, Asanuma M: Neuron-astrocyte interactions in Parkinson's disease. Cells 2020, 9:2623, https://doi.org/ 10.3390/cells9122623.
- MacMahon Copas AN, McComish SF, Fletcher JM, Caldwell MA: The pathogenesis of Parkinson's disease: a complex interplay between astrocytes, microglia, and T lymphocytes? Front Neurol 2021, 12, 666737, https://doi.org/10.3389/ fneur 2021 666737
- 7. Di Benedetto G, Burgaletto C, Bellanca CM, Munafò A, Bernardini R. Cantarella G: Role of microglia and astrocytes in

- Alzheimer's disease: from neuroinflammation to Ca<sup>2+</sup> homeostasis dysregulation. *Cells* 2022, 11:2728, https://doi.org/10.3390/cells11172728.
- Glass CK, Saijo K, Winner B, Marchetto MC, Gage FH: Mechanisms underlying inflammation in neurodegeneration. Cell 2010, 140:918–934, https://doi.org/10.1016/j.cell.2010.02.016.
- Strope TA, Birky CJ, Wilkins HM: The role of bioenergetics in neurodegeneration. Int J Mol Sci 2022, 23:9212, https://doi.org/ 10.3390/ijms23169212.
- Rangaraju V, Calloway N, Ryan TA: Activity-Driven local ATP synthesis is required for synaptic function. *Cell* 2014, 156: 825–835, https://doi.org/10.1016/j.cell.2013.12.042.
- Engl E, Attwell D: Non-signalling energy use in the brain. *J Physiol* 2015, 593:3417–3429, https://doi.org/10.1113/jphysiol.2014.282517.
- Aldana Bl: Microglia-specific metabolic changes in neurodegeneration. J Mol Biol 2019, 431:1830–1842, https://doi.org/ 10.1016/j.jmb.2019.03.006.
- Bernier L-P, York EM, MacVicar BA: Immunometabolism in the brain: how metabolism shapes microglial function. *Trends Neurosci* 2020, 43:854–869, https://doi.org/10.1016/ j.tins.2020.08.008.
- Chausse B, Malorny N, Lewen A, Poschet G, Berndt N, Kann O:
   Metabolic flexibility ensures proper neuronal network function in moderate neuroinflammation. Sci Rep 2024, 14, 14405, https://doi.org/10.1038/s41598-024-64872-1.

The metabolic reprogramming occurring in proinflammatory microglia, investigated by RNA-seq, molecular profiling and electrophysiology recordings, is accompanied by a decrease in the glucose:lactate ratio in the surrounding tissues. Computational simulations of a neuron energy model suggest that the variations in the extracellular glucose:lactate ratio could compromise neuronal fitness.

- 15. Cunnane SC, Trushina E, Morland C, Prigione A, Casadesus G, Andrews ZB, Beal MF, Bergersen LH, Brinton RD, de la Monte S, Eckert A, Harvey J, Jeggo R, Jhamandas JH, Kann O, la Cour CM, Martin WF, Mithieux G, Moreira PI, Murphy MP, Nave K-A, Nuriel T, Oliet SHR, Saudou F, Mattson MP, Swerdlow RH, Millan MJ: Brain energy rescue: an emerging therapeutic concept for neurodegenerative disorders of ageing. Nat Rev Drug Discov 2020, 19:609–633, https://doi.org/10.1038/s41573-020-0072-x.
- Wellstead P, Cloutier M: Modelling and simulation of brain energy metabolism: energy and Parkinson's disease. In Systems Biology of Parkinson's disease. Edited by Wellstead P, Cloutier M, New York, NY: Springer; 2012:19–38.
- Cloutier M, Bolger FB, Lowry JP, Wellstead P: An integrative dynamic model of brain energy metabolism using in vivo neurochemical measurements. *J Comput Neurosci* 2009, 27:391-414, https://doi.org/10.1007/s10827-009-0152-8.
- Cloutier M, Wellstead P: Dynamic modelling of protein and oxidative metabolisms simulates the pathogenesis of Parkinson's disease. *IET Syst Biol* 2012, 6:65–72, https:// doi.org/10.1049/iet-syb.2011.0075.
- Somersalo E, Cheng Y, Calvetti D: The metabolism of neurons and astrocytes through mathematical models. Ann Biomed Eng 2012, 40:2328–2344, https://doi.org/10.1007/s10439-012-0643-7
- Lloret-Villas A, Varusai T, Juty N, Laibe C, Le Novère N,
   Hermjakob H, Chelliah V: The impact of mathematical modeling in understanding the mechanisms underlying neurodegeneration: evolving dimensions and future directions. CPT Pharmacometrics Syst Pharmacol 2017, 6(2): 73–86, https://doi.org/10.1002/psp4.12155.

This comprehensive review has collected not less than 89 mathematical models for neurodegeneration, listing their main features and comparing their scope, to draw a general picture of the cellular and molecular mechanisms at play in the pathogenesis of AD and PD.

 Bakshi S, Chelliah V, Chen C, van der Graaf PH: Mathematical biology models of Parkinson's disease. CPT Pharmacometrics Syst Pharmacol 2019, 8(2):77–86, https://doi.org/10.1002/ psp4.12362.

- 22. Woo J, Cho H, Seol Y, Kim SH, Park C, Yousefian-Jazi A, Hyeon SJ, Lee J, Ryu H: Power failure of mitochondria and oxidative stress in neurodegeneration and its computational models. Antioxidants 2021, 10:229, https://doi.org/10.3390/ antiox10020229.
- Surmeier DJ, Obeso JA, Halliday GM: Selective neuronal vulnerability in Parkinson disease. Nat Rev Neurosci 2017, 18: 101-113, https://doi.org/10.1038/nrn.2016.178
- 24. Gonzalez-Rodriguez P, Zampese E, Surmeier DJ: Chapter 3 selective neuronal vulnerability in Parkinson's disease. In Progress in brain research. Vol. 252 of recent Advances in Parkinson's disease. Edited by Björklund A, Cenci MA, Elsevier; 2020:61–89, https://doi.org/10.1016/bs.pbr.2020.02.005.
- 25. Francis F, García MR, Middleton RH: A single compartment model of pacemaking in dissasociated Substantia nigra neurons. J Comput Neurosci 2013, 35:295-316, https://doi.org/ 10.1007/s10827-013-0453-9.
- Pissadaki E, Bolam JP: The energy cost of action potential propagation in dopamine neurons: clues to susceptibility in Parkinson's disease. Front Comput Neurosci 2013, 7:13.
- Chan CS, Gertler TS, Surmeier DJ: Calcium homeostasis, selective vulnerability and Parkinson's disease. Trends Neurosci 2009, 32:249-256, https://doi.org/10.1016/j.tins.2009.01.006
- Denton RM: Regulation of mitochondrial dehydrogenases by **calcium ions**. *Biochim Biophys Acta Bioenerg* 2009, **1787**: 1309–1316, https://doi.org/10.1016/j.bbabio.2009.01.005.
- 29. Griffiths EJ, Rutter GA: Mitochondrial calcium as a key regulator of mitochondrial ATP production in mammalian cells. Biochim Biophys Acta Bioenerg 2009, 1787:1324-1333, https:// doi.org/10.1016/j.bbabio.2009.01.019.
- 30. Zampese E, Wokosin DL, Gonzalez-Rodriguez P, Guzman JN, Tkatch T, Kondapalli J, Surmeier WC, D'Alessandro KB, De Stefani D, Rizzuto R, lino M, Molkentin JD, Chandel NS, Schumacker PT, Surmeier DJ: Ca2+ channels couple spiking to mitochondrial metabolism in substantia nigra dopaminergic neurons. Sci Adv 2022, 8, eabp8701, https://doi.org/ 10.1126/sciadv.abp8701.
- 31. del Arco A, González-Moreno L, Pérez-Liébana I, Juaristi I, \* González-Sánchez P, Contreras L, Pardo B, Satrúostegui J: Regulation of neuronal energy metabolism by calcium: role of MCU and Aralar/malate-aspartate shuttle. *Biochim Biophys Acta Mol Cell Res* 2023, **1870**, 119468, https://doi.org/10.1016/ i.bbamcr.2023.119468.

This paper is one of the most recent reviews about the regulatory role of  ${\rm Ca}^{2+}$  on mitochondrial bioenergetics, emphasising the role of  ${\rm Ca}^{2+}$  influx to mitochondria through the MCU and the  ${\rm Ca}^{2+}$  regulation of AGC/MAS.

- Soman S, Keatinge M, Moein M, Da Costa M, Mortiboys H, Skupin A, Sugunan S, Bazala M, Kuznicki J, Bandmann O: Inhibition of the mitochondrial calcium uniporter rescues dopaminergic neurons in pink1 zebrafish. Eur J Neurosci 2017, 45:528-535, https://doi.org/10.1111/ejn.13473
- Guzman JN, Sanchez-Padilla J, Wokosin D, Kondapalli J, Ilijic E, Schumacker PT, Surmeier DJ: **Oxidant stress evoked by** pacemaking in dopaminergic neurons is attenuated by DJ-1. Nature 2010, 468:696-700, https://doi.org/10.1038/nature09536.
- 34. Zorov DB, Juhaszova M, Sollott SJ: Mitochondrial reactive oxygen species (ROS) and ROS-induced ROS release. *Physiol Rev* 2014, **94**:909–950, https://doi.org/10.1152/ physrev.00026.2013.
- Piccirillo S, Magi S, Preziuso A, Serfilippi T, Cerqueni G Orciani M, Amoroso S, Lariccia V: The Hidden notes of redox balance in neurodegenerative diseases. Antioxidants 2022, 11: 1456, https://doi.org/10.3390/antiox11081456
- 36. Angelova PR, Abramov AY: Interplay of mitochondrial calcium signalling and reactive oxygen species production in the brain. *Biochem Soc Trans* 2024, **52**:1939–1946, https://doi.org/ 10.1042/BST20240261.
- Muddapu VR, Chakravarthy VS: Influence of energy deficiency on the subcellular processes of Substantia Nigra Pars Compacta cell for understanding Parkinsonian

neurodegeneration. Sci Rep 2021, 11:1754, https://doi.org/ 10.1038/s41598-021-81185-9.

This comprehensive model supports the validity of the energetic deficiency hypothesis by showing that the remodelling of multiple pathways (including Ca2+ signalling) observed in PD can be triggered by hypoxia and hypoglycaemia.

- 38. Post MR, Lieberman OJ, Mosharov EV: Can interactions between α-synuclein, dopamine and calcium explain selective neurodegeneration in Parkinson's disease? Front Neurosci 2018, 12:161, https://doi.org/10.3389/fnins.2018.00161
- 39. Burbulla LF, Song P, Mazzulli JR, Zampese E, Wong YC, Jeon S. Santos DP, Blanz J, Obermaier CD, Strojny C, Savas JN, Kiskinis E, Zhuang X, Krüger R, Surmeier DJ, Krainc D: **Dopa**mine oxidation mediates mitochondrial and lysosomal dysfunction in Parkinson's disease. Science 2017, 357: 1255-1261, https://doi.org/10.1126/science.aam9080
- 40. Poliquin PO, Chen J, Cloutier M, Trudeau L-E, Jolicoeur M: Metabolomics and in-silico analysis reveal critical energy deregulations in animal models of Parkinson's disease. PLoS One 2013, 8, e69146, https://doi.org/10.1371/ journal.pone.0069146.
- 41. Qi H, Li L, Shuai J: Optimal microdomain crosstalk between endoplasmic reticulum and mitochondria for Ca<sup>2+</sup> oscillations. *Sci Rep* 2015, **5**:7984, https://doi.org/10.1038/srep07984.
- Lim D, Dematteis G, Tapella L, Genazzani AA, Calì T, Brini M, Verkhratsky A: Ca<sup>2+</sup> handling at the mitochondria-ER contact sites in neurodegeneration. *Cell Calcium* 2021, **98**, 102453, https://doi.org/10.1016/j.ceca.2021.102453.
- LaFerla FM, Green KN, Oddo S: Intracellular amyloid-β in Alzheimer's disease. Nat Rev Neurosci 2007, 8:499–509, https://doi.org/10.1038/nrn2168.
- 44. Demuro A, Parker I, Stutzmann GE: Calcium signaling and amyloid toxicity in alzheimer disease. J Biol Chem 2010, 285: 12463-12468, https://doi.org/10.1074/jbc.R109.080895
- 45. Calvo-Rodriguez M, Bacskai BJ: Mitochondria and calcium in Alzheimer's disease: from cell signaling to neuronal cell death. Trends Neurosci 2021, 44:136-151, https://doi.org/10.1016/ j.tins.2020.10.004.
- 46. LaFerla FM: Calcium dvshomeostasis and intracellular signalling in Alzheimer's disease. Nat Rev Neurosci 2002, 3: 862-872, https://doi.org/10.1038/nrn960.
- 47. Pimplikar SW, Nixon RA, Robakis NK, Shen J, Tsai L-H: Amyloid-independent mechanisms in Alzheimer's disease pathogenesis. J Neurosci 2010, 30:14946-14954, https://doi.org/ 10.1523/JNEUROSCI.4305-10.2010.
- 48. Cheung K-H, Mei L, Mak D-OD, Hayashi I, Iwatsubo T, Kang DE, Foskett JK: Gain-of-Function enhancement of IP3 receptor modal gating by familial Alzheimer's Disease-Linked presenilin mutants in human cells and mouse neurons. Sci Signal 2010, 3:ra22, https://doi.org/10.1126/scisignal.2000818. ra22.
- Jensen L, Bultynck G, Luyten T, Amijee H, Bootman M, Roderick HL: Alzheimer's disease-associated peptide  $A\beta_{42}$ mobilizes ER Ca<sup>2+</sup> via InsP<sub>3</sub>R-dependent and -independent mechanisms. *Front Mol Neurosci* 2013, **6**:36.
- 50. Demuro A, Parker I: Cytotoxicity of intracellular  $A\beta_{42}$  amyloid oligomers involves  $Ca^{2+}$  release from the endoplasmic reticulum by stimulated production of inositol trisphosphate. JNeurosci 2013, 33:3824-3833, https://doi.org/10.1523/JNEUR-OSCI.4367-12.2013.
- 51. Toglia P, Ullah G: The gain-of-function enhancement of IP3receptor channel gating by familial Alzheimer's diseaselinked presenilin mutants increases the open probability of mitochondrial permeability transition pore. Cell Calcium 2016, 60:13-24, https://doi.org/10.1016/j.ceca.2016.05.002
- 52. Calvo-Rodriguez M, Kharitonova EK, Bacskai BJ: Therapeutic strategies to target calcium dysregulation in Alzheimer's disease. Cells 2020, 9:2513, https://doi.org/10.3390/
- Cortassa S, Aon MA, Marbán E, Winslow RL, O'Rourke B: An integrated model of cardiac mitochondrial energy

- metabolism and calcium dynamics. *Biophys J* 2003, **84**: 2734–2755, https://doi.org/10.1016/S0006-3495(03)75079-6.
- Cortassa S, Aon MA, Winslow RL, O'Rourke B: A mitochondrial oscillator dependent on reactive oxygen species. Biophys J 2004, 87:2060–2073, https://doi.org/10.1529/ biophysj.104.041749.
- Toglia P, Cheung K-H, Mak D-OD, Ullah G: Impaired mitochondrial function due to familial Alzheimer's diseasecausing presentlins mutants via Ca<sup>2+</sup> disruptions. Cell Calcium 2016, 59:240–250, https://doi.org/10.1016/ i.ceca.2016.02.013.
- Toglia P, Demuro A, Mak D-OD, Ullah G: Data-driven modeling of mitochondrial dysfunction in Alzheimer's disease. Cell Calcium 2018, 76:23–35, https://doi.org/10.1016/ i.ceca.2018.09.003.
- 57. Pensalfini A, Umar AR, Glabe C, Parker I, Ullah G, Demuro A:
   \* Intracellular injection of brain extracts from Alzheimer's disease patients triggers unregulated Ca<sup>2+</sup> release from intracellular stores that hinders cellular bioenergetics. *Cells* 2022, 11:3630, https://doi.org/10.3390/cells11323630

11:3630, https://doi.org/10.3390/cells11223630. Experimental Ca<sup>2+</sup> traces obtained in *Xenopus* oocytes upon injection of brain extracts from AD patients are used as input for a kinetic model of mitochondrial bioenergetics. Model predictions indicate that the altered Ca<sup>2+</sup> signals can induce anomalous ATP and ROS traces.

- Mak D-OD, Cheung K-H, Toglia P, Foskett JK, Ullah G: Analyzing and quantifying the gain-of-function enhancement of IP<sub>3</sub> receptor gating by familial Alzheimer's disease-causing mutants in presenilins. PLoS Comput Biol 2015, 11, e1004529, https://doi.org/10.1371/journal.pcbi.1004529.
- Latulippe J, Lotito D, Murby D: A mathematical model for the effects of amyloid beta on intracellular calcium. PLoS One 2018, 13, e0202503, https://doi.org/10.1371/ journal.pone.0202503.
- Minicucci J, Alfond M, Demuro A, Gerberry D, Latulippe J: Quantifying the dose-dependent impact of intracellular amyloid beta in a mathematical model of calcium regulation in xenopus oocyte. PLoS One 2021, 16, e0246116, https://doi.org/ 10.1371/journal.pone.0246116.
- 61. Farina S, Voorsluijs V, Fixemer S, Bouvier DS, Claus S, Ellisman MH, Bordas SPA, Skupin A: **Mechanistic multiscale**

- modelling of energy metabolism in human astrocytes reveals the impact of morphology changes in Alzheimer's Disease. *PLoS Comput Biol* 2023, **19**, e1011464, https://doi.org/10.1371/journal.pcbi.1011464.
- Hasel P, Aisenberg WH, Bennett FC, Liddelow SA: Molecular and metabolic heterogeneity of astrocytes and microglia. *Cell Metabol* 2023, 35:555–570, https://doi.org/10.1016/j.cmet.2023.03.006.
- Voorsluijs V, Avanzini F, Falasco G, Esposito M, Skupin A: Calcium oscillations optimize the energetic efficiency of mitochondrial metabolism. iScience 2024, 27, 109078, https:// doi.org/10.1016/j.isci.2024.109078.

This recent research paper combining nonquilibrium thermodynamics and kinetic modelling highlights the boosting effect of Ca<sup>2+</sup> on the energetic efficiency of mitochondrial metabolism in astrocytes.

- Strubbe-Rivera JO, Chen J, West BA, Parent KN, Wei G-W, Bazil JN: Modeling the effects of calcium overload on mitochondrial ultrastructural remodeling. Appl Sci 2021, 11:2071, https://doi.org/10.3390/app11052071.
- Kornick K, Bogner B, Sutter L, Das M: Population dynamics of mitochondria in cells: a minimal mathematical model. Front Physiol 2019, 7:146, https://doi.org/10.3389/fphy.2019.00146.
- Holt KB, Winter J, Manley S, Koslover EF: Spatiotemporal modeling of mitochondrial network architecture. PRX Life 2024, 2, 043002, https://doi.org/ 10.1103/PRXLife.2.043002.
- 67. Nair SS, Muddapu VR, Chakravarthy VS: A multiscale, \*\* systems-level, neuropharmacological model of cortico-basal ganglia system for arm reaching under normal, parkinsonian, and levodopa medication conditions. Front Comput Neurosci 2022, 15, 756881, https://doi.org/10.3389/fncom.2021.756881. doi:10.3389/fncom.2021.756881.

This multiscale framework of high translational value simulates motor symptoms (evaluated through a simulated arm reaching task) based on signalling network connecting different parts of the brain (cortico-basal ganglia model). The extracellular release of dopamine, predicted with a comprehensive SNc neuron model [37], is transmitted as an input signal to the basal ganglia. The ability of the model to determine the optimal levodopa dose in dependence on the state of the patient offers a potential clinical application.